

# *Laboratory Connectivity and Integration*

APHL Annual Meeting  
& First State Environmental Laboratory Conference  
Jacksonville June 4, 2007  
Rex Astles, PhD  
Acting Chief, LSDB, CDC



# OUTLINE

- Roots of the NLS and Recent Progress
- Case Studies
  - Washington Foundation for Healthcare Quality
  - Minnesota
  - Nebraska
- Progress with Antimicrobial Susceptibility Testing
  - Michigan
  - Current Multistate Consortium
- Discussion



# The Old Paradigm

- A loose association of public health (state, county and city), hospital, and independent laboratories throughout the country.



# Role of Laboratories

*“Provide information for decision making”*

## Private Labs

- Diagnostic testing
- Some reference testing
- Medical management
- Focus = Individual health

## Public Labs

- Some diagnostic testing
- Reference testing
- Surveillance and monitoring
- Focus = Public health

*Interdependent Network*

**Improve the Public's Health**



# A National Laboratory System

Linking public health, clinical, veterinary, food safety, and environmental laboratories to create seamless systems within each state for public health surveillance and laboratory support and improvement is the urgent mission of the National Laboratory System initiative

**Critical point:** the NLS depends upon strong State Public Health Laboratory Systems.

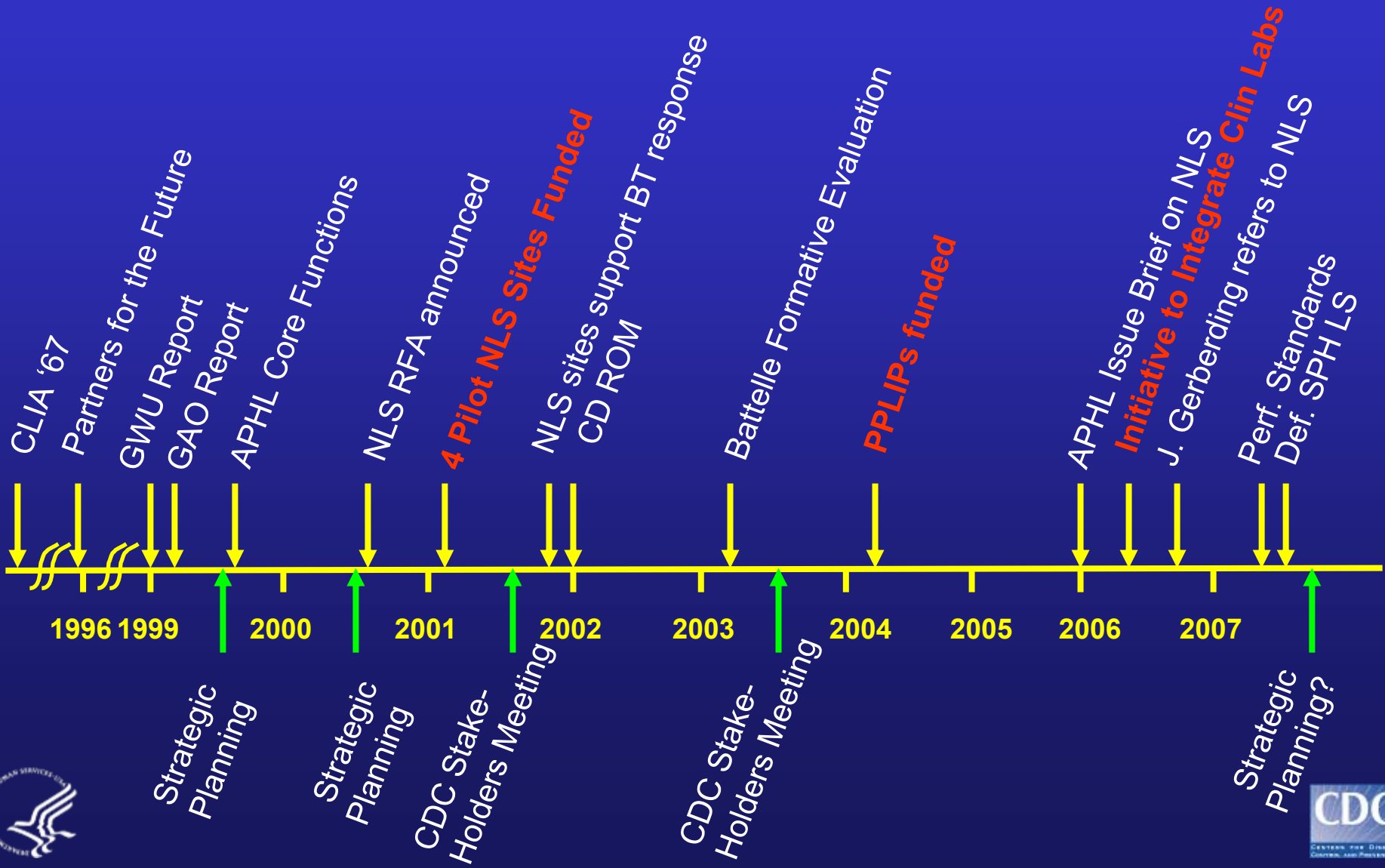


# What is the “State-Level Public Health Laboratory System?”

- *...More than the state public health laboratory*
  - All public, private, and voluntary entities that contribute to public health laboratory practice in the state
  - A network of entities with differing roles, relationships, and interactions



# NLS Developmental Timeline



# CDC's Global Health Protection Network

**FY04**

INTERNATIONAL

DOMESTIC

**FY04**

Quarantine & Border Health Stations

Field Epidemiology / Laboratory Training Programs

CDC Field Stations

International Business Connectivity

DOD Laboratories

New Global Disease Detection & Response Sites

International Rapid Response Teams

New International Laboratory Response Network

Sites **FY 07**

**Global Health Protection Network**

DATA EXCHANGE

**BioSense & Biointelligence Center**

**National Laboratory System**

National Clinical Lab Orders

DoD/VA Dx & Rx Records

Biowatch Data

Drug Sales

Clinical Care

US hospitals and clinics in sentinel cities

Disease Reporting

National Surveillance Systems

**FY 07**





# Look How Far We've Come

## APHL Survey, Summer 2001

	YES	NO	TOT.
Started new activities to improve clinical testing	14	21	35
Lab Advisory Committee	9	26	35
Newsletter for at least some Clinical Micro Labs	18	17	35
Have a BT Liaison	16	19	35
Regional Agreements w Other PHLs	15	20	35
Contact Clin. Labs to Assure Surveillance of Dz's	19	15	34
*3 employed by SPHL; 16 empl. by state epi. program	3/16*		



# Timely Opportunities

- Bioterrorism – Focal Area C
  - “Develop a plan to improve working relationships and communication between Level A (clinical ) laboratories and Level B/C laboratories, (i.e. Laboratory Response Network laboratories) as well as other public health officials.”
- Threat of Chemical Terrorism
- Emerging Threats
- OIG Report
- OSCAR Database
- CDC Reorganization



# System Components

*“What Gets Measured Gets Done”*

- Measurables
  - Core Functions
  - Healthy People 2010
  - COTPER Performance Goals
  - Performance Standards



# System Components (cont)

- Tools
  - Laboratory Program Advisors
  - National Center for PH Laboratory Leadership
  - National Laboratory Database
  - Core Functions
  - Performance Standards
- Extrapolations from “lessons learned”
  - <http://www.aphl.org/programs/LSS/partnership/Pages/default.aspx>



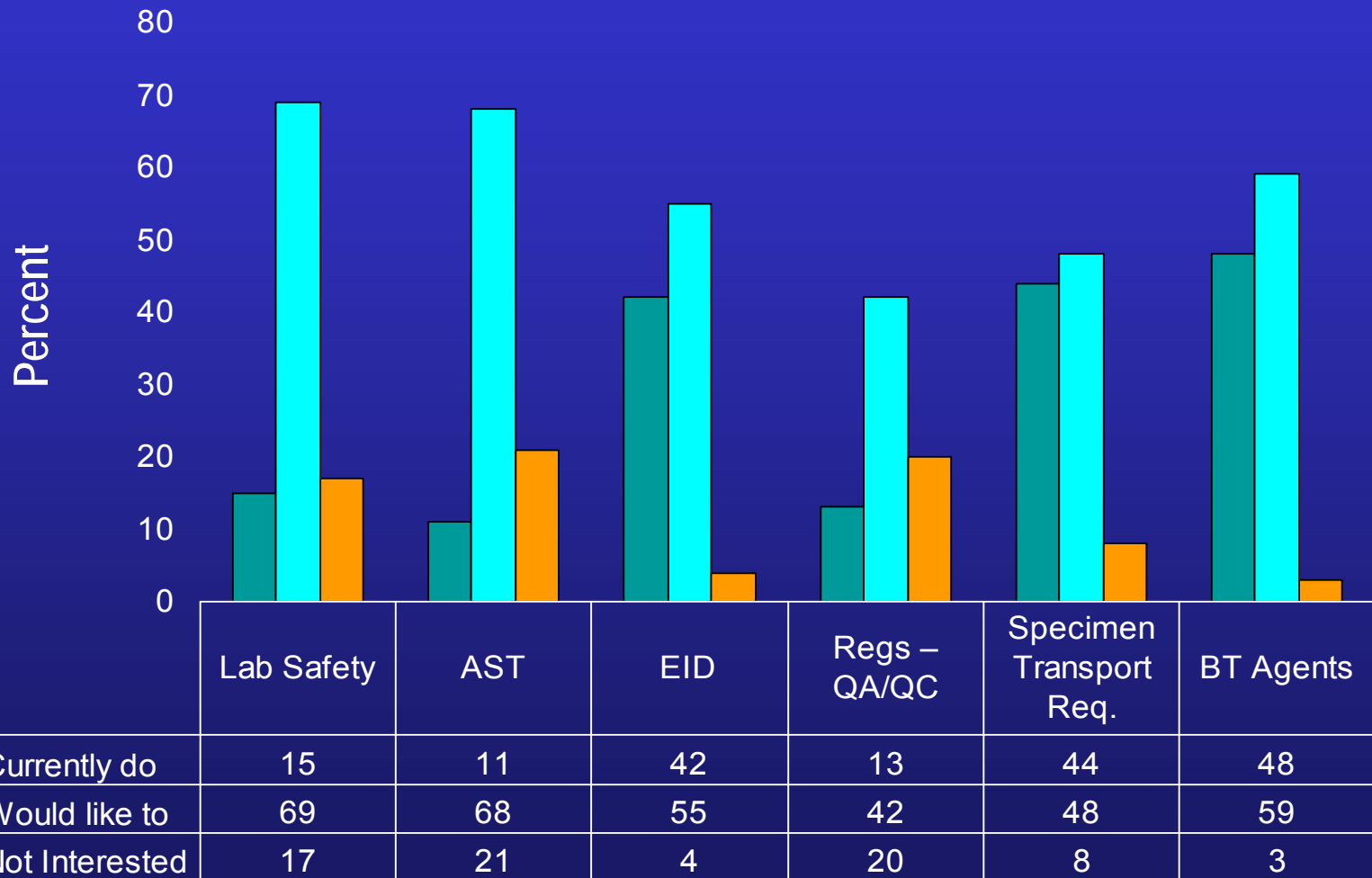
# Reasons CL Does Not Consult with SPHL

## - Battelle Formative Evaluation of the NLS Initiative

- 61% - Inability to quickly locate a point of contact
- 44% - Different hours of operation
- 19% - Not an appropriate source for some information
- 13% - Lack of confidence in SPHL expertise
- 10% - Concern about regulatory intervention
- 8% - Concern about interference in testing methods



# Clinical Lab Interest in Collaboration – Specific Topics



# PPLIP Activities

## Information Technology

- Connecticut
- Iowa
- Nebraska
- Rhode Island

## Communication

- Arkansas
- North Dakota
- Michigan

## Environmental Issues

- Minnesota
- Wisconsin

## Surveillance

- Massachusetts



# Defining the System

## Definition of a State Public Health Laboratory System

Association of Public Health Laboratories

June 2007

The State Public Health Laboratory System (SPH Laboratory System) consists of all the participants in public health testing, including those who initiate testing and those who ultimately use the test results. The SPH Laboratory System is part of the larger state public health system. The System includes individuals, organizations and agencies that are involved in assuring that laboratory data support the 10 Essential Services of Public Health. The concepts of a SPH Laboratory System are also embodied in the APHL Core Functions and Capabilities of State Public Health Laboratories. These documents are available on the APHL website at [www.aphl.org](http://www.aphl.org). Within the SPH Laboratory System are primary stakeholders who are directly involved in creating and using laboratory data. Additional stakeholders include those who are concerned with complementary Essential Services, such as Training and Education and Public Health Related Research. A successful National Laboratory System is dependent on the creation of fully integrated and coordinated networks in every state. The goals of the National Laboratory System are to support voluntary, interdependent partnerships of clinical, environmental, agricultural and veterinary laboratories through public-private collaboration, for assurance of quality laboratory services and public health surveillance.

The SPH Laboratory System should assure that:

1. public health threats are detected and intervention is timely
2. stakeholders are appropriately informed of potential threats
3. reportable conditions are monitored in a comprehensive statewide system
4. specimens and isolates for public health testing are sufficient to provide comprehensive public health surveillance and response
5. public health laboratory data are transmitted to appropriate state and federal agencies responsible for disease surveillance and control.

The state public health laboratory (SPHL) has a leadership role in developing and promoting the SPH Laboratory System through active collaboration with stakeholders, including epidemiologists; first responders; environmental professionals in water, food and air surveillance activities; private clinical and environmental laboratories; and local public health laboratories. The SPHL provides leadership to assure that essential and state-of-the-art laboratory services are provided and that clinical laboratories that perform public health testing on reportable infectious diseases submit results to the public health surveillance system using national testing guidelines. To provide leadership, the SPHL monitors essential components of the SPH Laboratory System, such as completeness of reporting and accuracy of laboratory testing results. The SPHL also assures that accurate results are reported in a manner that is appropriate and sufficiently timely for effective public health response. An effective SPH Laboratory System requires proactive leadership by the SPHL to monitor public health testing processes by clinical and environmental in-state laboratories. To assure that the SPH Laboratory System is effective, the SPHL should at a minimum:

1. maintain an integrated information system that includes all stakeholders that rely on accurate laboratory data
2. employ a full-time public health laboratory system coordinator
3. create a standing public health laboratory advisory committee
4. provide an interactive website or other electronic system to maintain regular communication channels for system partners.

*This document was developed by a subcommittee of the APHL Laboratory Systems & Standards Committee. It was adopted by the APHL Board on May 24, 2007.*



Association of Public Health Laboratories

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# Reaching Out Beyond Public Health: Strategies for Success

A **full-time** employee (preferably someone with commercial laboratory experience) to serve as **liaison** between the PHL and laboratory partners—the single most important resource on this list.

**Technology to enable rapid communication** between the PHL and its laboratory partners.

Resources to bring PHL staff together with laboratory partners for **face-to-face meetings, conferences and/or hands-on training workshops** of interest to laboratory partners.

A **database** containing information about all of a state or local jurisdiction's laboratory assets and the expertise to manage it.

A **Web site** designed for laboratory partners.

**Marketing capabilities** to explain what the PHL does, requirements for disease reporting, the benefits of participation in a state laboratory network and more.

A **laboratory advisory committee** comprised of stakeholders committed to common goals.

From APHL white paper "Building A National Laboratory System" March 2006  
[http://www.aphl.org/about\\_aphl/products\\_and\\_publications/Documents/lab\\_systems\\_3-06.pdf](http://www.aphl.org/about_aphl/products_and_publications/Documents/lab_systems_3-06.pdf)





Foundation for Health Care Quality

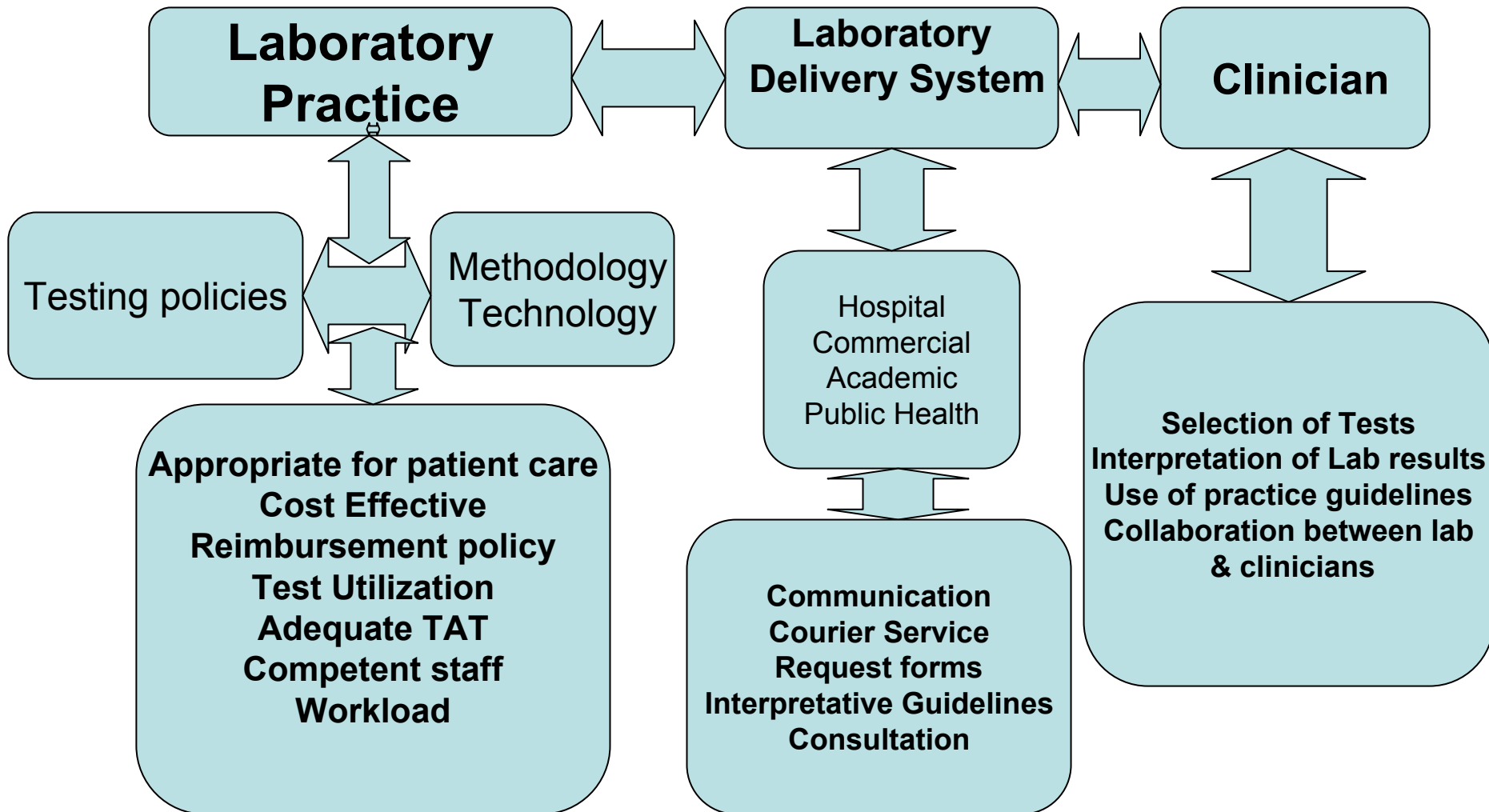
# **CLINICAL LABORATORY INITIATIVE**

JON M. COUNTS, DR.PH, MPH

10/12/2006



# Foundation for Health Care Quality



# PREVIOUS STUDIES

## 2000-2006

- Assessment/Improvement of AST
- Evaluation of Laboratory Delivery System
- Clinician Utilization of Laboratory Practice Guidelines



# SUMMARY OF IMPROVEMENTS IN AST LAB PRACTICE

- Acquisition of CSLI lab practice standards
- Performance in Case Studies
- Development/change in lab testing policies
- Use of Referral Laboratories by small micro labs



# LABORATORY DELIVERY SYSTEM

## Good to Excellent

- Lab accessibility for physicians (by phone, internet, email)
- Consultation with either a board-certified MD or PhD lab director
- Readability of reports
- Laboratory reports (format, content, usefulness)



# LABORATORY DELIVERY SYSTEMG-

## Good – Excellent

- Reliability of lab's courier service
- The information provided by lab on collection and submission of specimens/cultures
- Range of services performed by referral lab
- Willingness to accommodate special requests



# LABORATORY DELIVERY SYSTEM

## (Fair – Very Poor)

- Number and type of services provided on nights and weekends
- Quality of testing substandard
- Quality of technical consultation provided by lab
- Completion of request forms was onerous
- Lack of interpretative guidelines with reports





# LABORATORY DELIVERY SYSTEM

## (Fair – Very Poor)

- Turn-around time of tests
- Range of esoteric tests performed on-site
- Lab accessibility (physical location) for patients
- Failure to notify physician of critical test results



# CLINICIAN STUDIES

- Utilization of laboratory practice guidelines by primary care and infectious disease physicians (approx 5,000)
- Utilization of computerized physician order entry (CPOE)
- ID physician recommendations concerning antimicrobial testing and reporting- to inform CLAC guidelines
- Assessment of microbiology services provided by laboratory delivery system



# LABORATORY PRACTICE GUIDELINES

- Moderate awareness of CDC guidelines
- Low awareness of DOH/CLAC guidelines, but when aware, both were used for
  - Diagnosis
  - Testing
  - Communication with physicians
- Guidelines seen as difficult to use, complex, not helpful, not readily accessible
- Guidelines should be integrated into CPOE



# PROPOSED STUDIES

## 2006-2009

- Assess the inter-laboratory variability of laboratory practice, policies and processes in clinical microbiology in **small community hospital laboratories**
- Study factors which influence management decision-making, establishment of laboratory practice, current policies, and processes
- Evaluate methodology used to improve laboratory practice, policy and process measures that overtime would promote "best practices" in small hospital laboratories.



# PROPOSED STUDIES

## 2006 - 2009

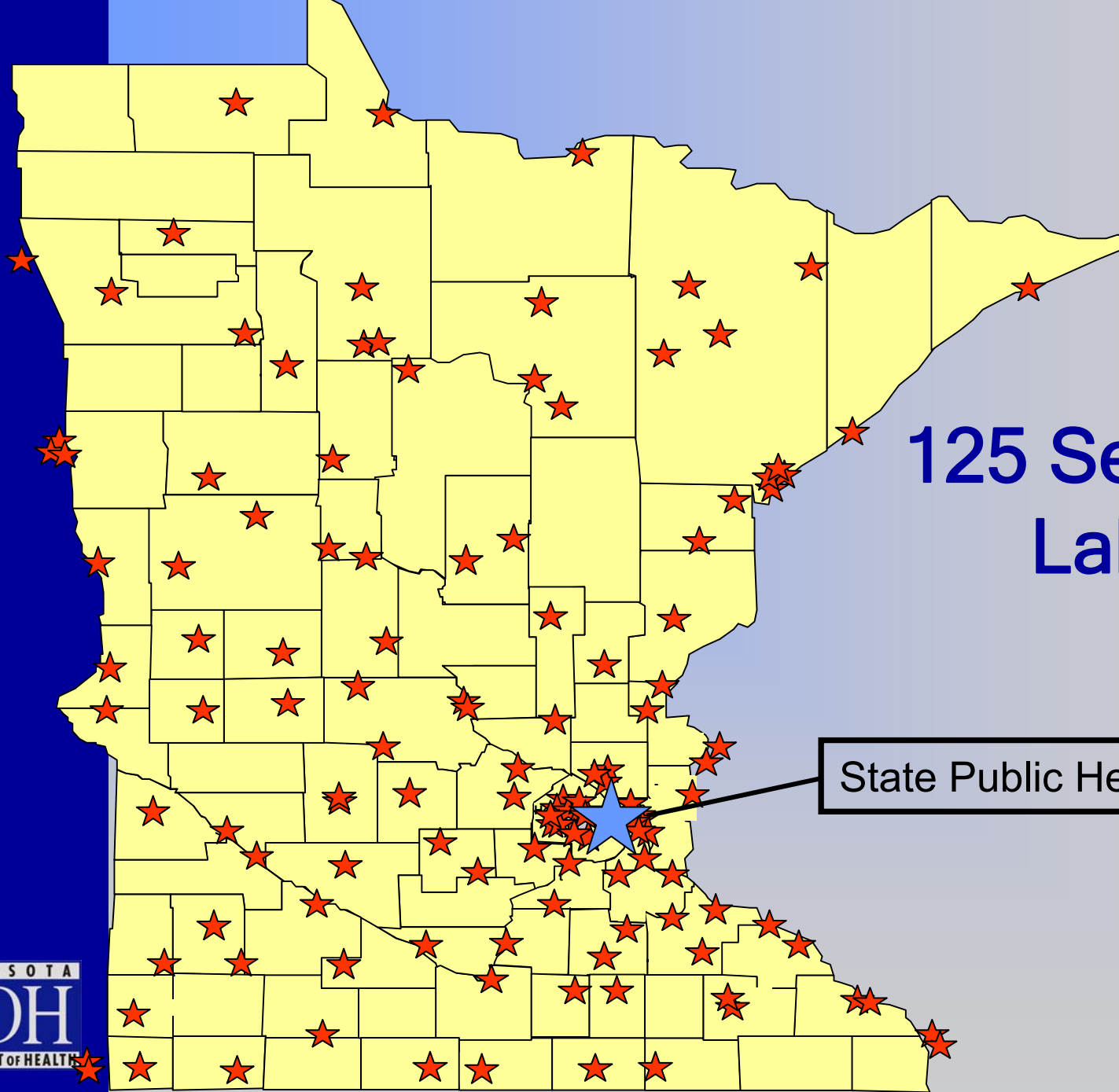
- Implement individualized quality management systems in Alaska, Oregon and Washington SPHL
- Improve their engagement and interaction with the laboratory delivery system in their state, including communication with the clinical laboratory community, quality of customer service and microbiology services provided by the SPHL
- Identify factors that impede their clinical laboratory community from adhering to voluntary national laboratory practice guidelines, reporting of results and submission of isolates and specimens to SPHL



# **“Re-visiting the Minnesota Laboratory System”**

**Paula M. Snippes**  
MT(ASCP)

**Program Advisor, MLS**



**125 Sentinel  
Labs**

State Public Health Lab

# Basic Components

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- Dedicated Program Advisor
- Recognizable system
- Robust communication
- Valuable products and programs
- Measurable benchmarks
- Supportive administration



# Dedicated Program Advisor

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- Full-time position
- Clinical lab background



# Recognizable System

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## Graphic Identifier



*An integrated network of public and private  
clinical laboratories working together  
to protect and improve the health of all  
Minnesotans*

# Recognizable System – MLS Website



**Minnesota Department of Health**  
Protecting, maintaining and improving the health of all Minnesotans

MDH HOME - [Minnesota Laboratory System](#)

**www.health.state.mn.us/mls**

**Minnesota Laboratory System**

- [MLS Home](#)
- [Education and Training](#)
- [Disease Specific Information](#)
- [MLS Lab Alerts](#)
- [Links](#)

**Public Health Laboratory**

- [Laboratory Home](#)
- [Environmental Laboratory](#)
- [Clinical Laboratory](#)
- [Newborn Screening](#)
- [Laboratory Emergency Preparedness](#)
- [Minnesota Laboratory System](#)
- [Laboratory Certification](#)

**MINNESOTA LABORATORY SYSTEM**  
A PRIVATE & PUBLIC COLLABORATION

*"An integrated network of public and private clinical laboratories working together to protect and improve the health of all Minnesotans"*

**About the Minnesota Laboratory System**

The Minnesota Laboratory System (MLS) is a statewide, voluntary network established by the Minnesota Department of Health - Public Health Laboratory to facilitate inter-laboratory communication, collaboration, and cooperation. Its members include public health and private clinical laboratories, as well as veterinary and agriculture laboratories, which serve Minnesota residents. A vital component of this system is electronic connectivity (fax and email) among all the members. In addition to providing a laboratory network essential for

**Hot Topics**

[Increase in Pertussis cases reported](#)  
Minnesota, North Dakota, Wisconsin have increased reporting rates of pertussis cases this year.

[West Nile Virus 2004](#)  
MDH-public health lab has new criteria for WNV testing in the current season. Also click for WNV links.

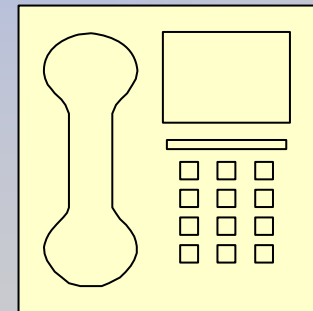
[MDH CLIA Certificate of Compliance](#)  
Now On-line



# Robust Communication

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- Robust database
- Blast email and fax capabilities
- Constant maintenance
- Listserve
- Website



# Robust Communication – MLS Lab Alerts



Categories of Lab Alerts:

## Laboratory **ALERT**

Conveys the highest level of importance; warrants immediate action, attention or response.

# Robust Communication – [MLS: e-LAB] - Listserv

MNLabSystem - Mail From: Ron.Jadwin@NorthMemorial.com

File Edit View Actions Tools Window Help

Close Reply Forward

Mail Properties Personalize

**From:** |  
**To:** "Minnesota Laboratory System" <mnlabssystem@listserv.health.state.mn.us>  
**BC:** |  
**Subject:** Re: [MLS:e-LAB] Screen plate for VRSA

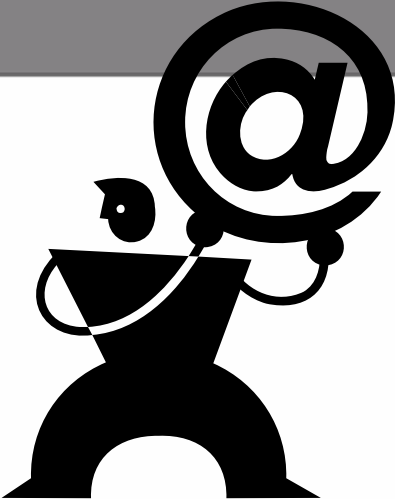
We set it up for all *S. aureus* isolates at the same time the Vitek card is set up. This works well into our workflow.

>>> | 9/1/04 12:48:09 PM >>>  
Are labs setting up the vancomycin screen plate on ALL Staph aureus isolates or only those that are positive for MRSA?  
r

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Confidentiality Notice: This email message, including any attachments, is for the sole use of the intended recipient(s) and may contain confidential and privileged information. Any unauthorized review, use, disclosure or distribution is prohibited. If you are not the intended recipient, please contact the sender by e-mail and destroy all copies of the original message or you may call the Queen of Peace Hospital Information Technology department at 952-758-8143.  
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To post a new subject: [mnlabssystem@listserv.health.state.mn.us](mailto:mnlabssystem@listserv.health.state.mn.us)  
To reply directly to the MDH Public Health Lab: [mnlabssystem@health.state.mn.us](mailto:mnlabssystem@health.state.mn.us)  
To add, remove or change your subscriber information: [mnlabssystem@health.state.mn.us](mailto:mnlabssystem@health.state.mn.us)  
Minnesota Laboratory System website: <http://www.health.state.mn.us/mls>

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The Minnesota Department of Health does not verify nor take responsibility for the accuracy of the statements posted on this listserv.



# Valuable Products and Programs

Education

Collaboration

## MLS Goals

- Enhance quality of microbiology practice
  - Antimicrobial susceptibility testing
  - Pathogen detection and identification
- Improve emergency preparedness
  - Bioterrorism and Chemical terrorism
  - Outbreak detection
- Provide resources/educational material
- Ensure communication and collaboration

Communication

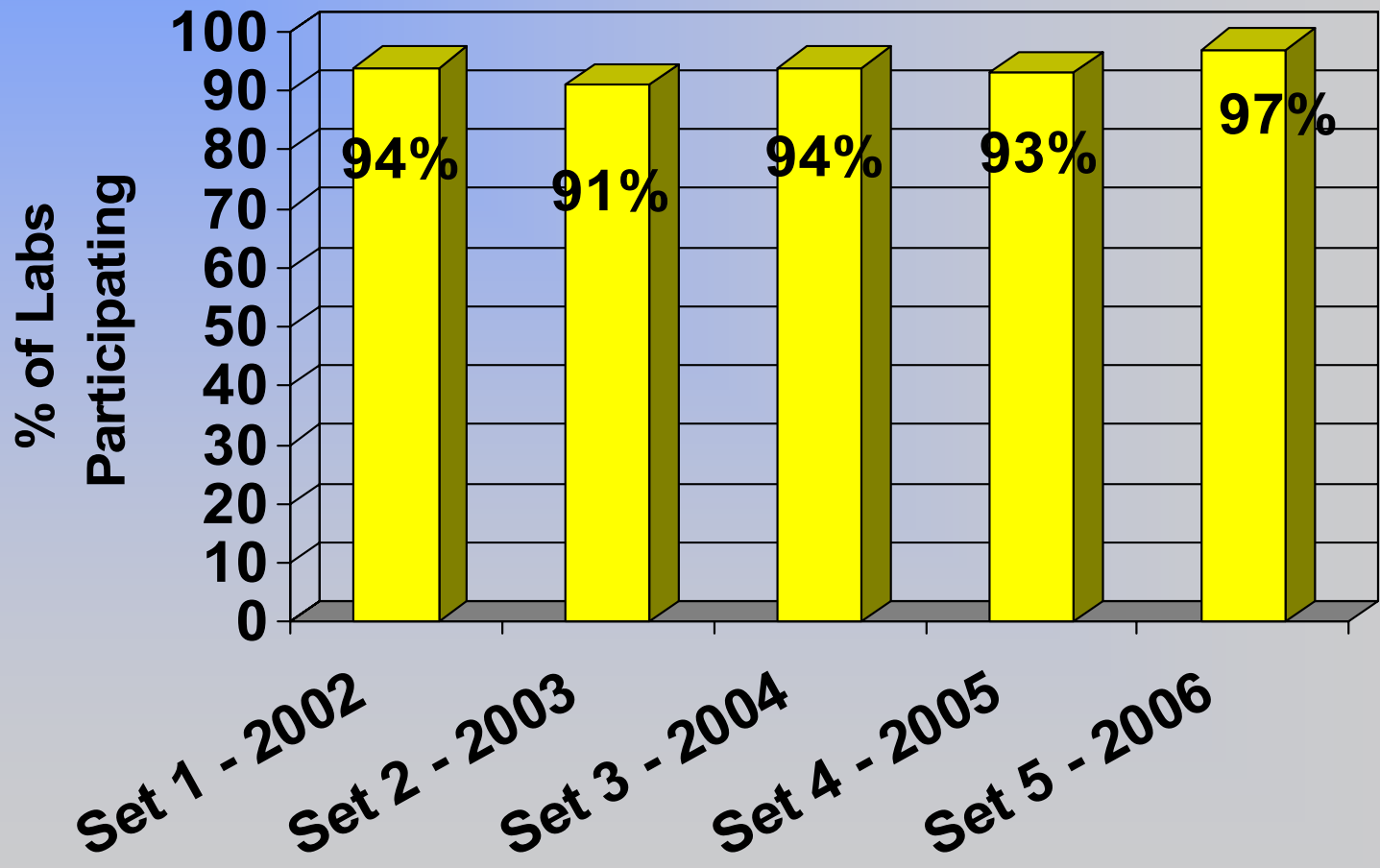
# Valuable Products and Programs

- 85 CLSI guidelines sent – 2006/07
- 313 participants MLS Regional Lab Conference – 2006
- 61 participants 2006 - BT Wet Workshops
- 135 BT/CT posters
- Challenge Set





# Valuable Products and Programs Challenge Set



# Valuable Products and Programs Challenge Set

## Goals

- Identify needs
- Monitor preparedness
- Assess practices/capabilities
- Provide educational resources
- Educate about diseases of public health importance

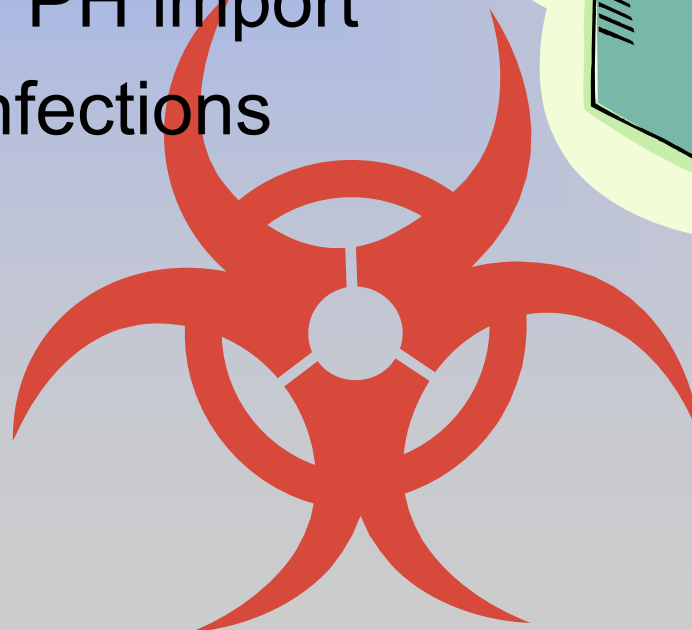


# Valuable Products and Programs Challenge Set

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## Organism Choices

- Terrorism-like agents
- Antibiotic resistance
- Diseases of PH import
- Emerging infections



# Valuable Products and Programs Challenge Set

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## Organisms (Set 5)

1. *Oligella ureolytica*
2. *Mycobacterium abscessus*
3. *Listeria monocytogenes*
4. Streptococcus group B (*S. agalactiae*)



# Valuable Products and Programs Challenge Set

## Findings:

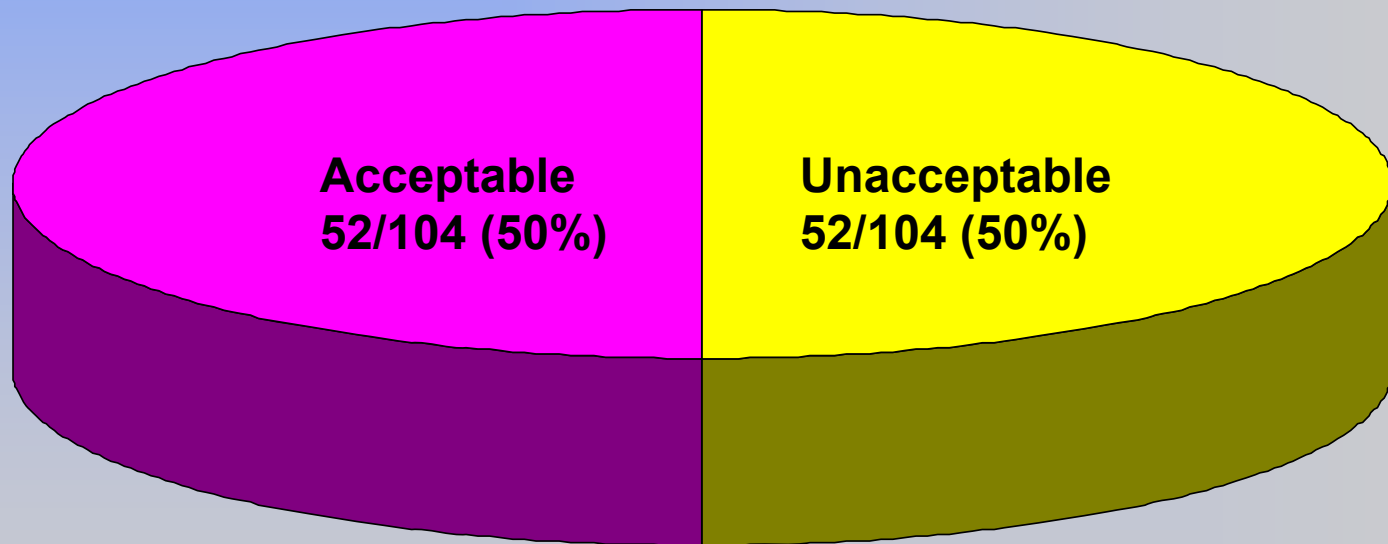
- Unusual gram-negative bacilli are difficult to ID
- Mycobacterium are not on the radar
- Changes in AST guidelines are a challenge
- Labs are interested - teleconference
  - 58 phone lines
  - 275 participants
  - 26 CDs



# Challenge Set Findings

*Brucella* surrogate (*O. ureolytica*)

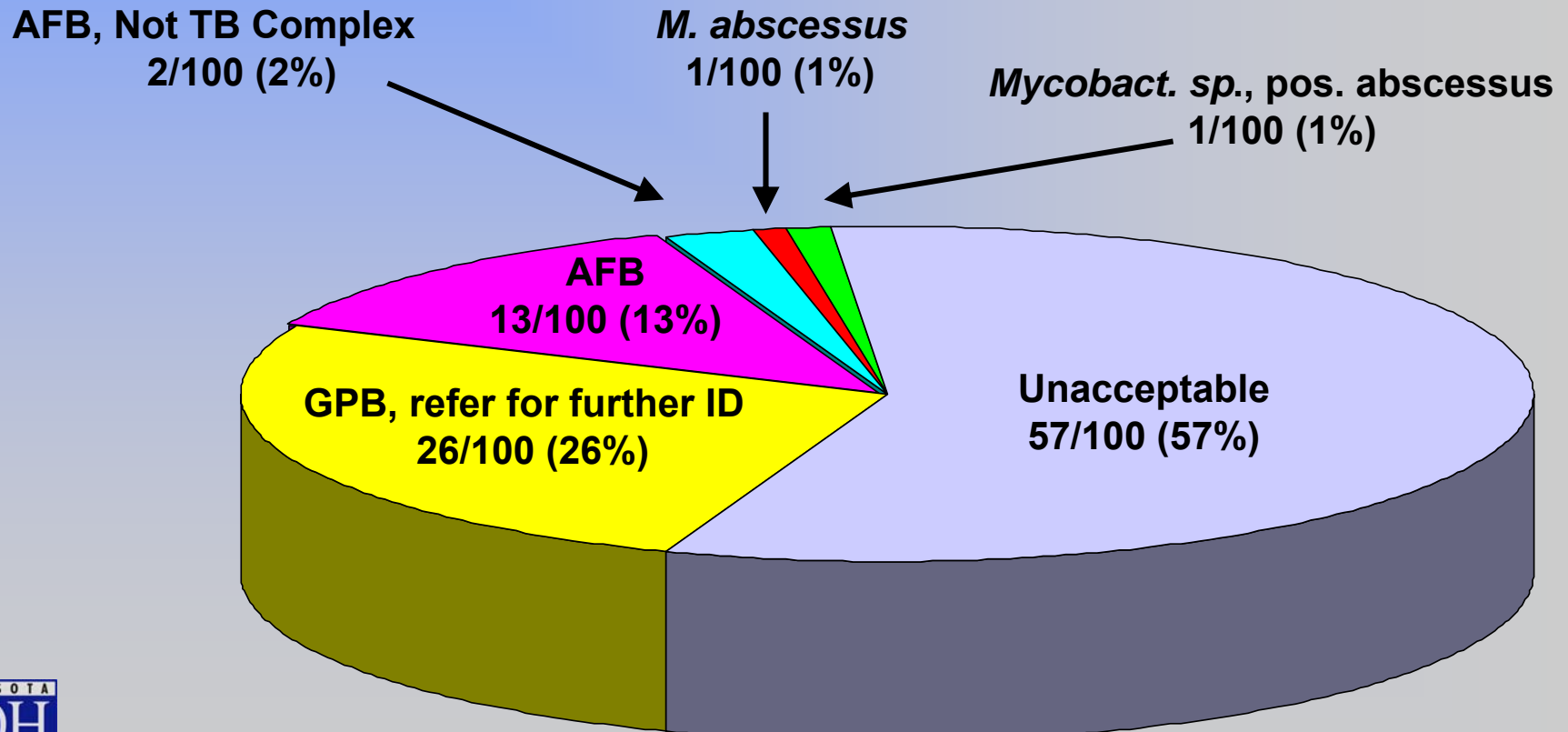
Identification (n=104)



# Challenge Set Findings

## *Mycobacterium abscessus*

Acceptable Answers 43/100 (43%)

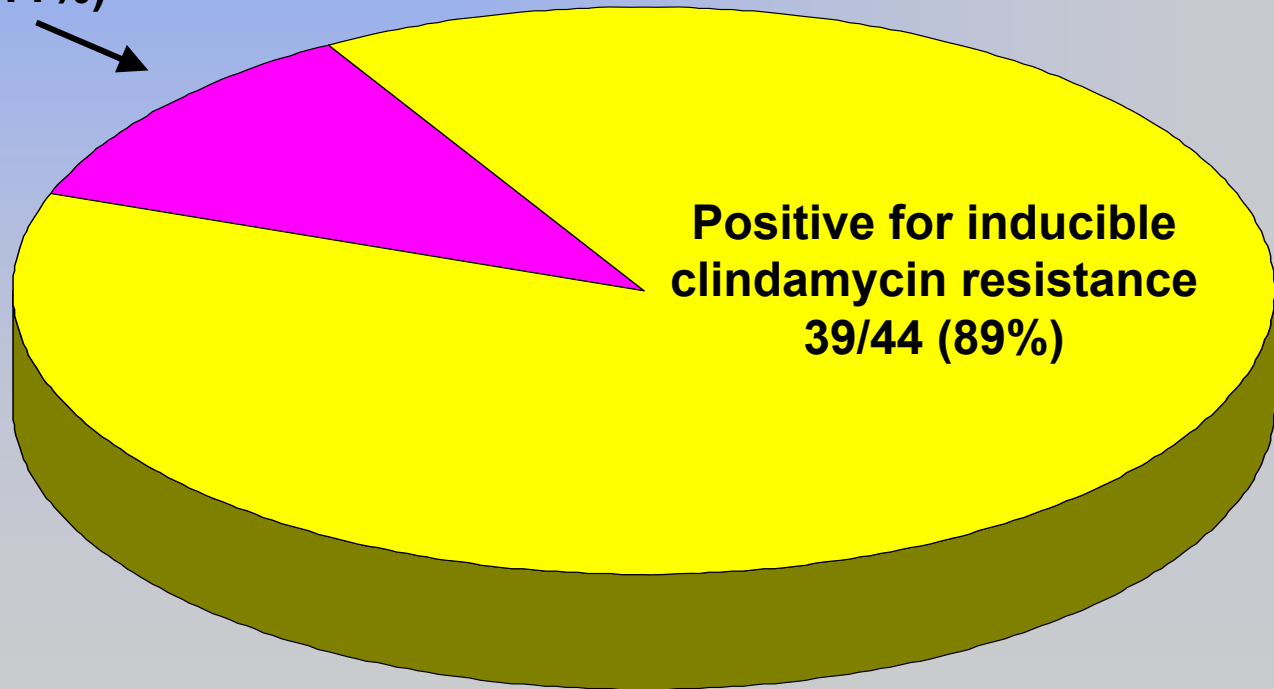


# Challenge Set Findings

## Group B Streptococcus (AST)

Performed D-zone Test (44/93 = 47%)

Negative for inducible  
clindamycin resistance  
5/44 (11%)



Positive for inducible  
clindamycin resistance  
39/44 (89%)



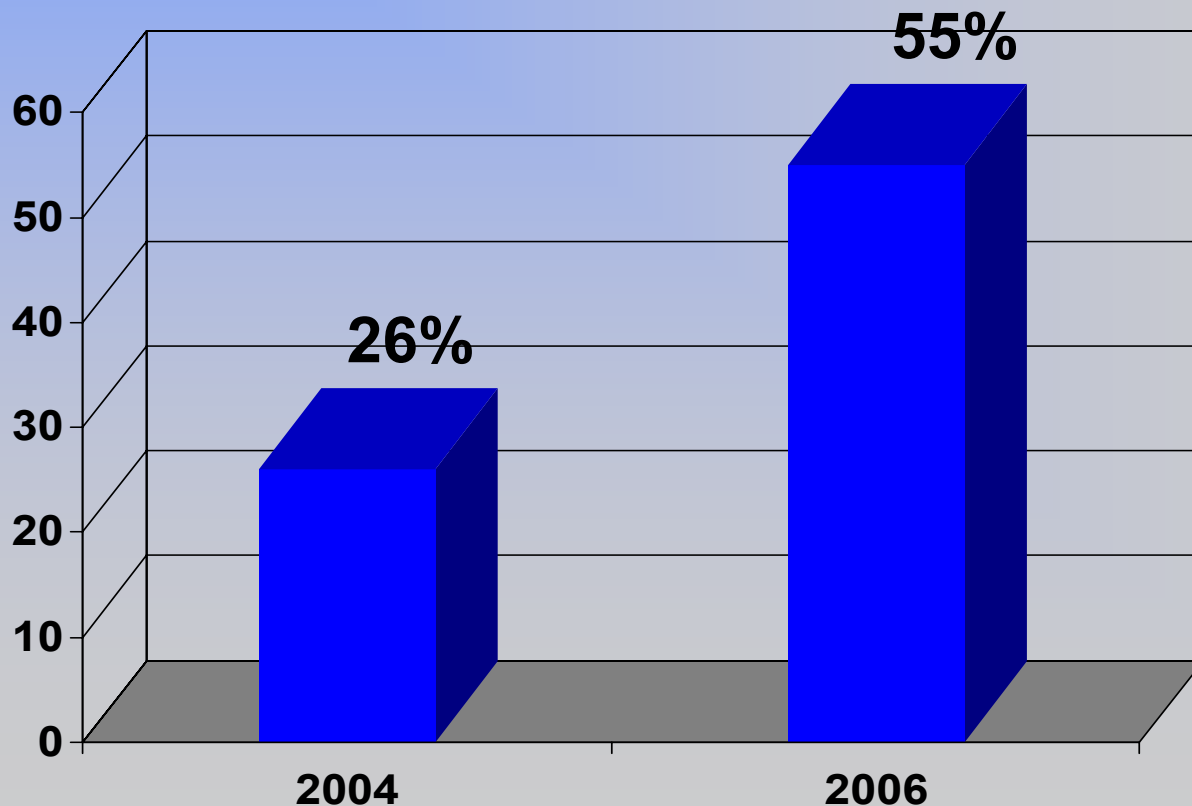
# Measurable Benchmarks Challenge Set

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- Staph aureus D-test
  - Direct comparison of two consecutive challenge sets indicated a 21% increase in the number of laboratories that perform the D-zone test.
- Group B strep

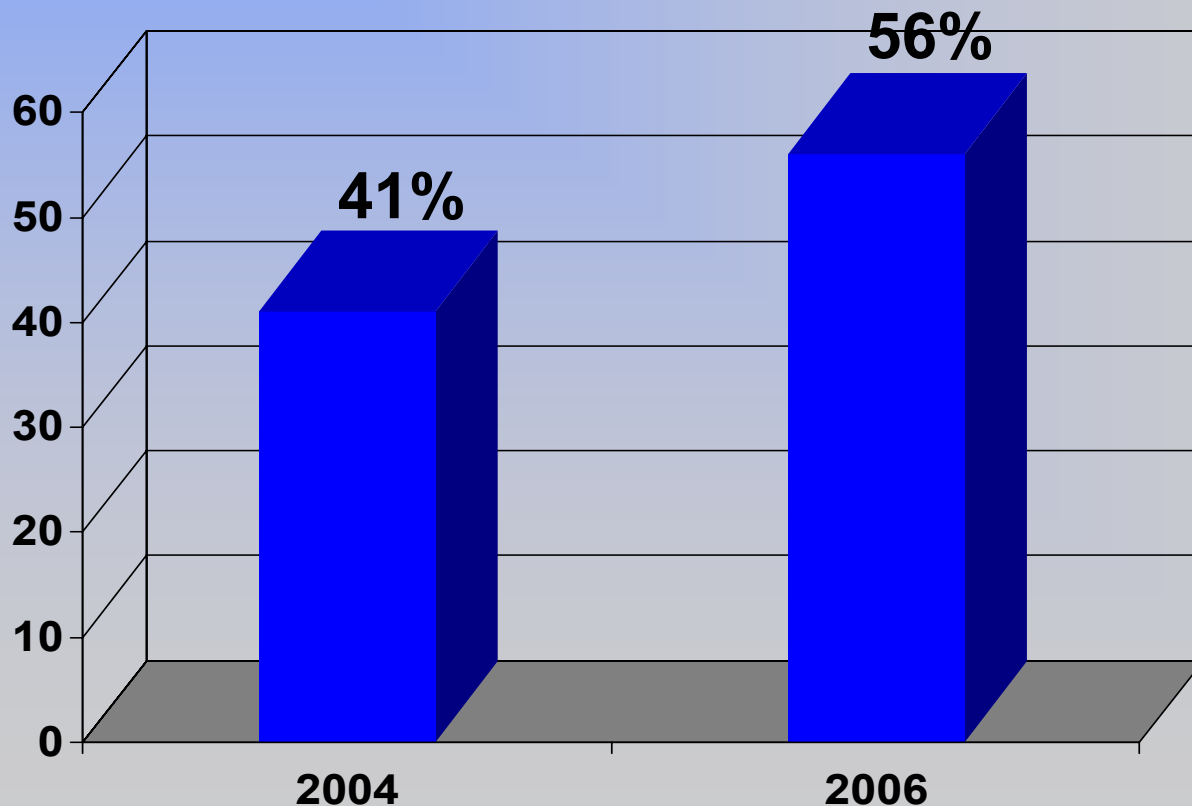
# Group B Streptococcus Disease Prevention

Penicillin allergy status information  
received by laboratory



# Group B Streptococcus Identification

Identification of ALL GBS in urine cultures of pregnant women



# Measurable Benchmarks Challenge Set

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## Challenge Set - Evaluation Comments

- “....sometimes leads to changes in procedures--good feedback.”
- “It helps educate people who do not have a strong microbiology background.”
- “It gives us an idea about our performance with comparison with other labs.”

# Supportive Administration

- Support: full-time position
- Resources
  - Funds
  - Staff
  - Tools



# Barriers

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- Resource Heavy
- Support: full-time position
- Resources
  - Funds
  - Staff
  - Tools



# Final Words

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- Change is slow
- Persistence is essential
- Impact is Positive





**THANK YOU**

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**Paula M. Snippes**

Program Advisor, Minnesota  
Laboratory System (MLS)

**651-201-5581**

[paula.snippes@health.state.mn.us](mailto:paula.snippes@health.state.mn.us)



# Building Blocks of the Nebraska Laboratory Network (NLN)

Steve Hinrichs, M.D., Director, NPHL

Tony Sambol, MA, Associate Director, NPHL

Josh Rowland, MBA, MT(ASCP), State Training  
Coordinator, NPHL

# Laboratory Demonstration Project (LDP)-Phase 1

- 2001-APHL
- Situational assessment
- Communication
  - Labs connected only by phone-no internet
- Large geographical area
  - “Distance” barriers for rapid testing
- Infrastructure
  - Loosely woven or non-existent with NPHL

# Phase 1

- Survey: site visits assessed BT knowledge and communication needs-identify sentinel labs
- Communication: possible satellite links and GIS
- Training: refine bioterrorism training material
- Product: CD-ROM with BT materials
- Laid groundwork for the future...

# LDP-Phase 2

- 2002-2003 FAC funding
- Designated hub labs within the 6 regional population centers
- Regional level-A (sentinel) training-didactic lectures
- Improved communication infrastructure
- Hired a State Training Coordinator
- Developed lab “buy-in” through needs assessment survey
- Further buy-in with site visits of all facilities

# Phase 2

- Continued “communication” enhancements
- Reinvigorated [www.nphl.org](http://www.nphl.org), NPHL Newsletter, ELR ...
- STATPack™ Ver 1.0
- Regional conferences and workshops
- Teleconferences - NLTN
- “The face of PH in NE”

# LDP-Phase 3

- 2004 PPLIP-CDC funding
- Integration with environmental, food, and veterinary diagnostic labs into NLN
- STATPack™ and cross training workshops
- Now have MOU



# To Date LDP Outcomes

- Enhanced opportunities to interact with NLN
- BT wet workshops offered since 2004, 38 of 45 labs
- Evidence that labs communicate with NPHL readily
- CT preparedness workshops well attended
- Too hard to quantify...but NLN better prepared future BT/CT/PH events

# 2006 Project

- DLS/CDC-Initiative to Integrate Private Laboratories into Public Health Testing
- Goal: assess/develop an antimicrobial susceptibility testing educational program in Nebraska
- Paul D. Fey, Ph.D. Associate Professor/Associate Director, NPHL
- Josh Rowland, MBA, MT(ASCP) State Training Coordinator



# Specific aims:

- Determine needs through personal interview and AST survey
- Develop and implement “hands on” wet labs and lectures
- Develop a long term consultation solution through consultative telemedicine-STATPack, 6 additional units in 3 years

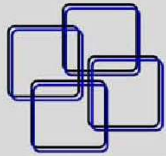
# STATPack™

- **Secure Telecommunications Application Terminal Package**
- Remote electronic consultation-telemedicine
- HIPAA compliant
- Video/camera images
  - Macro/microscopic
- Education/case studies
- Priority Levels
- 20 sites in NE (11 in OK, 9 in KS)



# STATPack™ Case Studies

- AST Case, February 2007
- Initial message to NE labs included AST results (Erythromycin R, Clindamycin S)
- Technique, interpretation, and methodology of the D-test (positive D-test shown) were discussed



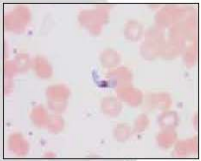
A.L. Fruhling<sup>1</sup>, M.L. Lund<sup>1</sup>, D.J. Rowland<sup>2</sup>, R.K. Noel<sup>2</sup>, B.K. Schweitzer<sup>2</sup>, J.J. Mathewson<sup>3</sup>, J.F. Murray<sup>3</sup>, R.E. Flahart<sup>4</sup>, S.E. Gabel<sup>4</sup>, S.A. Riley<sup>4</sup>, K.M. Weiss<sup>1</sup>, G.A. Hoff<sup>1</sup>, A.R. Sambol<sup>2</sup>, S.H. Hinrichs<sup>2</sup>.

<sup>1</sup>College of Information Science & Technology, University of Nebraska at Omaha, Omaha, NE, <sup>2</sup>Nebraska Public Health Laboratory, University of Nebraska Medical Center, Omaha, NE, <sup>3</sup>Oklahoma State Department of Health, Oklahoma City, OK, <sup>4</sup>Kansas Department of Health and Environment, Topeka, KS.

## Consultation

### Malarial

Images sent by St. Mary's Hospital in Enid, OK to consult with Oklahoma Public Health Laboratory one week after installation and training on STATPack. Determined to be *Plasmodium falciparum* based on morphological characteristics.



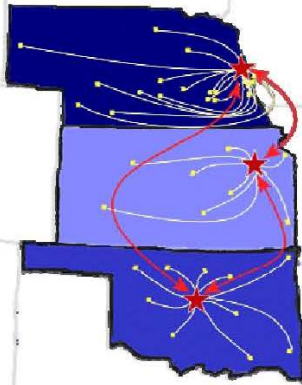
### Fungal

Images sent to the Nebraska Public Health Laboratory (NPHL) with a request for fungal consultation. Unable to rule-out *Stachybotrys* lymphadenitis. Recommendation was for the submitting laboratory to send the specimen to a reference laboratory for further characterization.



### Bacterial

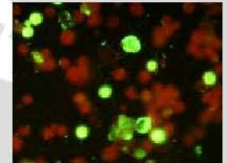
Images received at the NPHL for consultation. Catalase positive, non-hemolytic, non-motile, large GPR from a blood culture. Sentinel laboratory could not rule-out *Bacillus anthracis*. Morphology was not consistent with *Bacillus anthracis*.



## Education

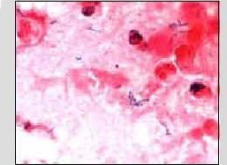
### VZV DFA

Six sentinel laboratories across Nebraska were trained to perform Varicella-Zoster Virus (VZV) Direct Fluorescent Antibody testing as a rule-out for Variola virus (smallpox). This picture represents a positive control slide that one laboratory sent to NPHL.



### Competency Training

A series of Gram-stained slides serve as a repository for clinical competency training and documentation of proficiency. This is used by a clinical hospital laboratory in Nebraska to satisfy their College of American Pathologist (CAP) gram stain competency requirement (MIC.21565).



### Case Study

One example of a STATPack case study was sent to laboratories in Nebraska. The initial STATPack message included antimicrobial susceptibility testing results (Erythromycin R, Clindamycin S). Technique, interpretation, and methodology of the D-test (positive D-test shown) were discussed in this exercise.



# National Laboratory System: Initiative to Integrate Private Laboratories into Public Health Testing

Laurina O. Williams, PhD, MPH  
Project Officer  
CDC/NCID/NCPDCID  
Division of Laboratory Systems



# National Laboratory System – CDC Staff

## *Initiative to Integrate Clinical Laboratories into Public Health Testing*

### Division of Laboratory Systems

Joe Boone, PhD  
Acting Director, DLS

John Ridderhof, DrPH  
Acting Deputy Director, DLS

Laurina Williams, PhD, MPH  
Project Officer

Rex Astles, PhD  
Project Officer

Pam Robinson  
Program Analyst

Jesse Holder  
Programmer

### Division of Healthcare Quality Promotion

Roberta Carey, PhD  
Branch Chief, ELB

Fred Tenover, PhD

Clifford McDonald, MD

Brandi Limbago, PhD

Jean Patel, PhD

Shalein Banerjei, PhD

### Consultants

Vanessa White, APHL

Jim Hidalgo, APHL

Rosemary Humes, APHL

Janet Hindler, PhD

NLTN



# National Laboratory System (NLS) Initiatives

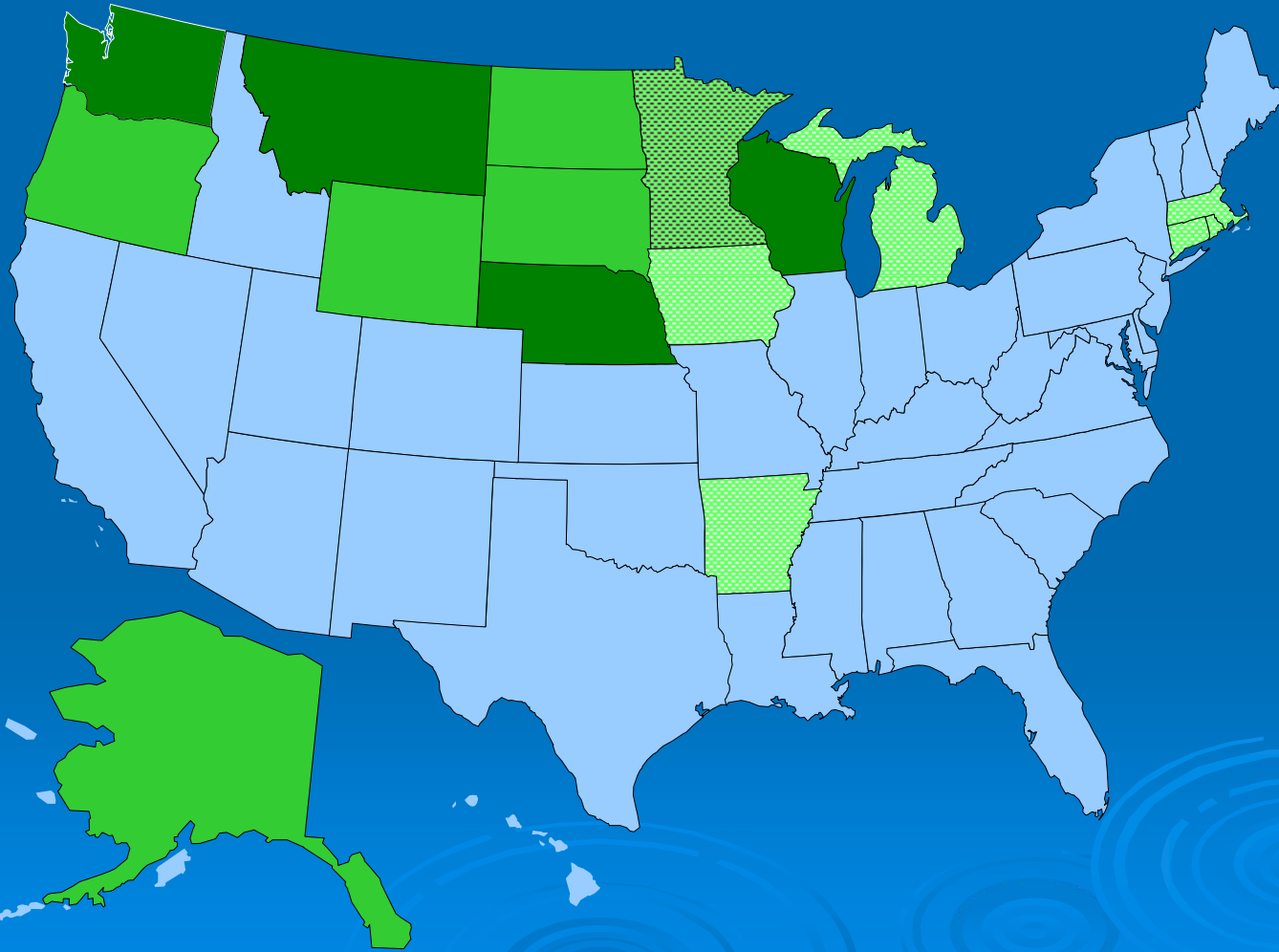
DLS is working with partners and through public-private laboratory integration projects to strengthen the NLS, an enhanced communication and collaboration network among public health and clinical laboratories to facilitate:

- High quality and timely public health laboratory testing
- Improved assessment of relevant laboratory practices
- Better detection of, response to, and tracking of public health threats
- An effective mechanism for making policy and adopting appropriate guidelines across states and regions
- Development of performance standards





# National Laboratory System *States Ever Included as NLS Projects*



# Current NLS Activities

## ➤ **AST Assessment**

- Montana (Northern Plains Consortium); Nebraska, Wisconsin
- 250 participants (full participation needed)
- Survey covers demographics, methods, guidelines, outcomes

## ➤ **STD-related Activities**

- Montana - Northern Plains Consortium
- Foundation for Healthcare Quality

## ➤ **General Clinical Microbiology Practices**

- Foundation for Healthcare Quality



# Current NLS Project Management

- **Montana – Northern Plains Consortium**  
Project Director – Anne Weber  
Project Supervisor – Susie Zanto  
Program Coordinator – Debbie Gibson  
AST Project manager – John LaRue
- **North Dakota:**  
Myra Kosse  
Eric Hieb  
Representative: Danita Hunke
- **Wyoming:**  
Rich Harris  
Representative: Jim McKinna
- **South Dakota:**  
Mike Smith  
Representative: Yvette Thomas
- **Wisconsin:**  
Project Directors: Steve Marshall and Carol Kirk
- **Nebraska:**  
Project Directors: Paul Fey and Josh Rowland
- **Foundation for Healthcare Quality:**  
Project Director: Jon Counts



# AST Assessment Activities

**AST Laboratory Practices Survey  
Consensus Process  
Data Analysis Consensus**

**Antibiogram Worksheet and Tools**

**Interventions**

**ASCP Course**

**Onsite Training and Consultation**

**STAT-Pak implementation (Nebraska)**



# Survey Development and Committees

## Methods-Chair:

Paul D. Fey  
Jean Patel  
Roberta Carey  
Fred Tenover

## Demographics-Chair

Jon M. Counts/ Rex Astles  
Anne Pollock  
Brandi Limbago

## Guidelines-Chair

Joni Wedig  
Clifford McDonald

## Outcomes-Chair

John LaRue  
Laurina Williams  
Pam Thompson  
Debbie Gibson  
Steve Marshall  
Susie Zanto

## Statistician

Shalein Banerjei



# Survey Development Process

Consensus Process – Modeled after CLSI Document Process  
(November 2006)

## Content Committees

- Demographics
- Methods
- Guidelines
- Outcomes

Committees composed questions

Consensus Conference Calls

CDC compiled “final” first draft

Statistical Review

- Types of questions vs. statistical power

Pilot testing by the Montana Consortium

Final Consensus Questionnaire (March 2007)

Individual States could add questions at end of survey



# The AST Questionnaire

## Antimicrobial Susceptibility Testing (AST) Questionnaire

### Cover Page:

Today's Date \_\_/\_\_/\_\_\_\_ (MM/DD/YYYY)

Please provide information about yourself:

Name:

---

Laboratory Name:

---

CLIA Number:

---

Street Address:

---

City: \_\_\_\_\_ State: \_\_\_\_\_ Zip

Code: \_\_\_\_\_

Telephone: \_(\_\_\_\_) \_\_\_\_\_ - \_\_\_\_\_

Email Address:

---

Additional information about yourself:

- Medical laboratory director
- Laboratory manager - administrative
- Technical consultant
- General supervisor
- Specialty supervisor
- Non-supervisory testing personnel
- Other, please specify: \_\_\_\_\_



53. Please complete the table by following the steps below.

**STEP 1.** Complete column 1 by putting an "X" in the box next to the organism if testing of any type is performed at your laboratory.

- For any organism you checked, complete Steps 2 and 3. Do not complete Steps 2 or 3 for organisms you did not check.

**STEP 2.** Complete column 2 by entering the estimated number of isolates tested for each organism.

**STEP 3.** Complete columns 3, 4, and 5 using the codes below to list screening and primary and secondary/confirmatory testing methods for each organism. (Please use capital letters to fill in the table.)

Rather than leave a space blank, use code 'A' if your laboratory doesn't perform a test method. If code 'W' is selected, please specify the procedure in column 6. If more than one comment is needed per organism, be sure to clarify which column number the comment addresses.

Organism	1 AST is performed on organism (Check Box)	2 Estimated Annual AST Volume for 2006	3 Screening test methods	4 Primary test methods	5 Secondary or confirmatory testing methods	6 Comments
<i>Enterococcus</i> spp.	<input type="checkbox"/>					
<i>Staphylococcus aureus</i>	<input type="checkbox"/>					
Coagulase negative <i>Staphylococcus</i>	<input type="checkbox"/>					
<i>Streptococcus pneumoniae</i>	<input type="checkbox"/>					
<i>Streptococcus agalactiae</i>	<input type="checkbox"/>					
<i>Enterobacteriaceae</i>	<input type="checkbox"/>					
<i>Haemophilus influenzae</i>	<input type="checkbox"/>					
<i>Pseudomonas aeruginosa</i>	<input type="checkbox"/>					
<i>P. aeruginosa</i> from cystic fibrosis	<input type="checkbox"/>					
Non- <i>Enterobacteriaceae</i>	<input type="checkbox"/>					
HACEK group <sup>†</sup>	<input type="checkbox"/>					
Gram positive anaerobes	<input type="checkbox"/>					
Gram negative anaerobes	<input type="checkbox"/>					

**List of Codes**

<b>A</b> =Do not perform this method with this organism	<b>I</b> =BD Phoenix	<b>P</b> =D-zone test for inducible clindamycin resistance
<b>B</b> =Oxacillin Screen agar (MHA+Salt)	<b>J</b> =Microscan Walkaway Rapid	<b>Q</b> =Cefoxitin disk test for MRSA
<b>C</b> =Vancomycin Screen agar (BHI agar)	<b>K</b> =Microscan Walkaway Conventional	<b>R</b> =Latex agglutination (PBP) testing method for MRSA
<b>D</b> =PCR	<b>L</b> =Microscan auto or touchscan	<b>S</b> =ESBL confirmation test with clavulanic acid (Disk Diffusion)
	<b>M</b> =Trek (Sensititre /Axis)	<b>T</b> =ESBL confirmation test with clavulanic acid (E-test)
<b>E</b> =E-test methodology	<b>N</b> =Vitek 2	<b>U</b> =ESBL confirmation test with clavulanic acid (automated testing)
<b>F</b> =Kirby Bauer disk diffusion	<b>O</b> =Vitek Legacy	<b>V</b> =β-lactamase production (e.g. nitrocephin disk)
<b>G</b> =Agar Dilution MIC methodology		<b>W</b> =Other testing procedure (please specify in Column 6.)
<b>H</b> =Broth microdilution MIC methodology (manual reading)		<b>X</b> =Unknown

<sup>†</sup>Includes *Acinetobacter*, *Bacteroides*, and *Xenorhabdium*

<sup>†</sup>Includes *Haemophilus* (not *H. influenzae*), *Actinobacillus*, *Cardiobacterium*, *Eikenella*, and *Kingella*





57. When testing a community-associated methicillin resistant *Staphylococcus aureus* (CA-MRSA) strain that is resistant to penicillin and oxacillin, which of the following antimicrobials would you report as resistant? (Check all that apply, even if you do not report them in your laboratory.)

- |  |                                       |
|--|---------------------------------------|
| <input type="checkbox"/> I don't know                | <input type="checkbox"/> Cefazolin    |
| <input type="checkbox"/> Ampicillin-sulbactam        | <input type="checkbox"/> Ceftriaxone  |
| <input type="checkbox"/> Amoxicillin-clavulanic Acid | <input type="checkbox"/> Imipenem     |
| <input type="checkbox"/> Erythromycin                | <input type="checkbox"/> Tetracycline |
| <input type="checkbox"/> Vancomycin                  |                                       |

62. Which of the following isolates, or presumptive isolates does your laboratory refer to a reference laboratory for additional testing/ confirmation? (Check all that apply.)

- We do not refer
- VISA or VRSA (vancomycin intermediate or resistant *Staphylococcus aureus*)
- Streptococcus pneumoniae* from a sterile site
- VRE (vancomycin resistant enterococci)
- MRSA (methicillin resistant *Staphylococcus aureus*)
- ESBL (extended spectrum beta-lactamase producers)

65. In your opinion was this survey: (Check all that apply.)

- Important
- Relevant
- Educational
- Appropriate
- None of the above

67. Were the questions clear?

- Yes                       No

If no, which questions were unclear? \_\_\_\_\_



# Survey Data Group

**John LaRue - Montana**

**Neil Squires, Programmer - Montana**

**Debbie Gibson - Montana**

**Susie Zanto - Montana**

**Bonnie Barnard - Montana**

**Kammy Johnson, Epidemiologist – Montana**

**Eric Hieb – North Dakota**

**Gale Stevens - Wyoming**

**Yvette Thomas – South Dakota**

**Chris Carlson – South Dakota**

**Steve Marshall – Wisconsin**

**Joni Wedig – Wisconsin**

**Dave Warshaer - Wisconsin**

**Paul Fey - Nebraska**

**Josh Rowland - Nebraska**

**Shalein Banerjee, Statistician – CDC/DHQP**

**Jesse Holder, Programmer – CDC/DLS**

**Laurina Williams – CDC/DLS**



# Survey Data Group Process

- **Consensus Process for Editing Instructions**
  - Reviewed each question – interpretation; validity
  - Statistical Review - “reasonableness” and analytical considerations
- **Period of Review by data group and by entire survey group**
- **Editing Rules**
  - Extra information
  - Conflicting information – in most cases requires verification by state
- **Programming by Montana Consortium**
  - Incorporating editing rules and skip patterns
  - Comment section by data entry personnel
  - Future web version - compatible with ACCESS and SEQUEL
  - Concatenating data



# Data Entry Rules for 2007 NLS-AST Questionnaire

## General Rules

Some of the problems encountered while entering data into the data base may require calls to the participating facilities. In order to simplify this process, do not make a call to a facility until the entire survey has been reviewed; then you can verify all of the necessary information at one time.

If a question is left blank, you may skip the question until you are able to verify the data. It is suggested that each data entry person keep a handwritten log designating the facility and of all of the questions that need data verification. When the verification call is made, the log may be used as a reference. This handwritten log will need to be saved in the event that it needs to be reviewed at a later date. An Excel spreadsheet has been created to input this data for safe keeping.

Please be sure that the “knowledge-based” questions (54-62 and either #48 or #51) all have some type of answer collected. We will need as many participants as possible to answer these questions in order to compute a knowledge index.

The following “general” guidelines have been developed in an attempt to address the majority of translation problems, and they may be used for all questions unless the data editing instructions for a question explicitly overrides these guidelines. If any of the guidelines are unclear, please contact Debbie at 406-444-5970 for further clarification.

1. The six digit accession number: The database will automatically enter the first two digits, which will be the FIPS code for each state. The data entry person will need to assign the remaining four digits, which should be a unique four digit laboratory identifier.
2. This same unique accession number will be used each year for the same laboratory. Each year in the preference section of the data base, you will change the year to the current year, keeping the accession number the same. This way you will easily be able to look up data from a certain facility, designated by year.
3. For all questions in which “**other**” is an option, the database will require a comment. If “**other**” is marked in a question and nothing is specified in the blank, please enter “**No Comment**”.
4. Some questions will require call for data verification. After the call is made, you may change the answer from ‘No Comment’ to the actual comment gathered on the phone call.

John LaRue, Montana



## Statewide Cummulative Antibiogram Data CY 2006

Gram Positive Organism	# of isolates, all sources	Penicillin	Oxacillin (cefoxitin disc)	Cefazolin	Erythromycin	Trimethoprim-sulfamethoxazole	Cefuroxime	Cefotaxime	Clindamycin	Ciprofloxacin	Levofloxacin	Vancocmycin	Tetracycline	Linezolid	Quinupristin-dalfopristin	# of Isolates, Urine Only	Tetracycline (urine isolates)	Nitrofurantoin (urine isolates)
		Percent susceptible (%S)															(%S)	
* <i>Enterococcus</i> spp. (see explanation)	304	98%								71%	65%	100%	40%	96%	ND	63	33%	100%
<i>E. faecalis</i>	1672	98%								64%	71%	99%	28%	96%	ND	323	32%	99%
<i>E. faecium</i>	126	32%									25%	74%	71%	95%	92%	20	65%	75%
<i>S. aureus</i>	5807	9%	60%	58%	46%	99%	75%		76%	67%	70%	100%	95%	99%	100%	173	91%	97%
<i>S. pneumoniae</i>	406	63%			63%	79%	84%	94%	46%		100%	100%	85%	100%				

Gram Negative Organism	# of isolates, all sources	Ampicillin	Cefazolin	Ceftazidime	Gentamicin	Trimethoprim-sulfamethoxazole	Ampicillin / sulbactam	Piperacillin / tazobactam	Cefuroxime	Cefotaxime	Ceftioxone	Cefepime	Ciprofloxacin	Levofloxacin	Imipenem	Meropenem	Aztreonam	Amikacin	Tobramycin	Tetracycline	# of Isolates, Urine Only	Ciprofloxacin (urine isolates)	Levofloxacin (urine isolates)	Norfloxacin	Tetracycline (urine isolates)	Nitrofurantoin
		Percent susceptible (%S)																				(%S)				
<i>E. coli</i>	11594	64%	94%	98%	95%	85%	72%	98%	92%	99%	99%	100%	93%	93%	100%	100%	99%	100%	96%	82%	3902	84%	84%	79%	71%	44%
<i>K. pneumoniae</i>	2079		97%	98%	99%	96%	89%	94%	91%	98%	98%	100%	98%	98%	100%	100%	98%	99%	99%	91%	665	78%	77%	74%	71%	31%
<i>P. mirabilis</i>	719	84%	95%	99%	95%	84%	91%	99%	97%	99%	99%	99%	89%	95%	98%	100%	98%	98%	95%		204	69%	65%	70%		
<i>P. aeruginosa</i>	1369			91%	83%			98%		12%	22%	89%	75%	80%	90%	95%	75%	95%	95%		237	35%	36%	44%		

# The A-cumulator Tool

Laboratory 300006's antibiogram is stored under **Lab6** worksheet in the **Labs 1-60** workbook (**Figure 4**).

**Figure 4.** *S. aureus* data for laboratory 300006 stored in **Lab6** worksheet.

Gram Positive Organism	# of Isolates, all sources	Penicillin	Oxacillin (cefoxitin disc)	Cefazolin	Erythromycin	Trimethoprim-sulfamethoxazole	Cefuroxime	Cefotaxime	Clindamycin	Ciprofloxacin	Levofloxacin	Vancocycin	Tetracycline	Linezolid	Quinupristin-dalfopristin	# of Isolates, Urine Only	Tetracycline (urine isolates)	Nitrofurantoin (urine isolates)
		<b>Percent susceptible (%S)</b>																
*Enterococcus spp. (see explanation)																		
<i>E. faecalis</i>	26	92%								85%		100%	12%			11	18%	100%
<i>E. faecium</i>	8	12%										38%	88%			1	0%	100%
<i>S. aureus</i>	117	12%	44%		32%	100%			82%	76%		100%	98%			9	88%	100%
<i>S. pneumoniae</i>	6	100%																

**What happens then?** The data from the **Labs 1-60** workbook is pulled into and distributed to the appropriate pathogen worksheets in the **A-cumulator** workbook. At the bottom of each of the pathogen worksheets, the aggregated % susceptibility (%S) for each antimicrobial is calculated.

J. LaRue, Montana



## Getting Started:

Before you begin copying and pasting antibiograms, here are some additional actions that you will want to accomplish.

1. **Rename the archived files:** Montana has chosen to rename all of the antibiograms that have been returned with the accession number of the laboratory. (**Figure 2**) To save space, you may wish to delete the instructions worksheet that has been returned with each antibiogram file.

**Figure 2.** Renamed antibiograms in archive file.

- **Transcribe hardcopy data** (if necessary): Hopefully the majority of the antibiograms that are returned to you are in electronic format however we are finding a significant number of laboratories are printing the worksheet, filling it out by hand, and faxing or mailing it back to us via snail-mail. It is for this reason that the “**template-Antibiogram\_wrkst07**” was included (**Figure 2**). The data entry person will need to manually transfer the hard-copy susceptibility information into the template worksheet and save it as the appropriate laboratory.
1. **Change laboratory numbers in pathogen worksheets:** Another change that will need to be made is with regard to the **A-cumulator** workbook. Each pathogen sheet aggregates the susceptibility data from all of the laboratories that submit. See the example of the *S. aureus* worksheet in **Figure 3**. Each state that uses this workbook will need to change the accession numbers in **Column A** on each pathogen worksheet so that they correlate with the accession numbers they have assigned the laboratories in their respective states.

Each row represents the data from one laboratory. In **Figure 3**, Row 9 contains the data for laboratory 300006. Laboratory 300006’s antibiogram is stored under **Lab6** worksheet in the **Labs 1-60** workbook (**Figure 4**).

John LaRue, Montana



# ASCP Course

## “The Clinical Laboratory Standards Institute and the Microbiology Laboratorian: Putting Guidelines Into Practice”

### I. Course Introduction

#### Case Studies

Case studies on using the CLSI susceptibility testing guidelines to solve common questions and problems in the microbiology laboratory

#### Section 1: Using Tables 1 and 1A

III. (Suggested groupings of U.S. FDA-approved antimicrobial agents that should be considered for routine testing and reporting on nonfastidious organisms by clinical microbiology laboratories)

#### A. Case 1. Site-specific reporting of susceptibility results

Case 2. Selective reporting of susceptibility results

Case 3. Susceptibility reporting by antibiotic class

Case 4. Organism-specific reporting of susceptibility results

Case 5. Susceptibility reporting by antibiotic class (2nd example)

#### Section Two: Using Tables 2A-2I

Case 6. Detecting and reporting inducible clindamycin resistance in staphylococci

Case 7. Detecting and reporting extended-spectrum beta-lactamase (ESBL) resistance in enterobacteriaceae

Case 8. The changing epidemiology of Methicillin-resistant *Staphylococcus aureus* (MRSA)

Case 9. Susceptibility testing of *Staphylococcus lugdunensis*

#### Section Three: Using Tables 4 (M2) and 8 (M7)

(Suggestions for verification of antimicrobial susceptibility test results and confirmation of organism identification)

Case 10. Detecting testing errors and unlikely or unusual susceptibility results

#### Section Four: Other susceptibility testing issues

Case 11. Susceptibility testing when there are no CLSI-approved methods or interpretation breakpoints

Case 12. Design and use of an institution antibiogram

Additional Resources and CLSI Documents

Some Questions from Laboratorians & Answers





# STATPack™

- **Secure Telecommunications Application Terminal Package**
- Remote laboratory consultation on a variety of specimens (bacteria, fungal, AST...)
- 20 units currently in Nebraska (6 more to be deployed in three years of CDC AST grant)
  - 11 in Oklahoma, 9 in Kansas
- [www.statpack.org](http://www.statpack.org)

Josh Rowland, Nebraska



# Project Reporting

## ➤ From each State Leader

- Monthly reports to Consortium Leader (Montana)
- Progress, Obstacles, Outcomes

## ➤ NLS Partners

- Quarterly Reports to CDC
  - Activities
  - Obstacles
  - Proposed Solutions
  - Progress toward Expected Outcomes



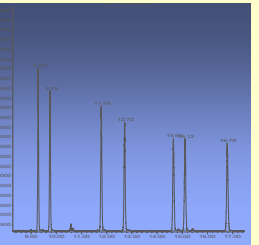
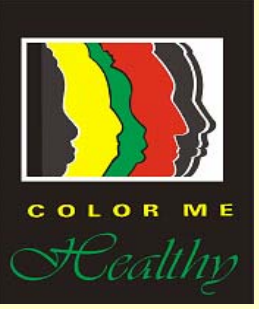
# Project Outcomes/Evaluation

- **NLS Partners in collaboration with CDC**
  - **Completion of Logic Models**
    - Ongoing
      - Annual
      - Overall Project
- **Overall project outcomes – CDC/NLS Mission**
  - **Improve communications and collaboration between clinical labs and public health laboratories**
    - Improve laboratory testing practices
    - Improve public health



# Vision: Integrated System





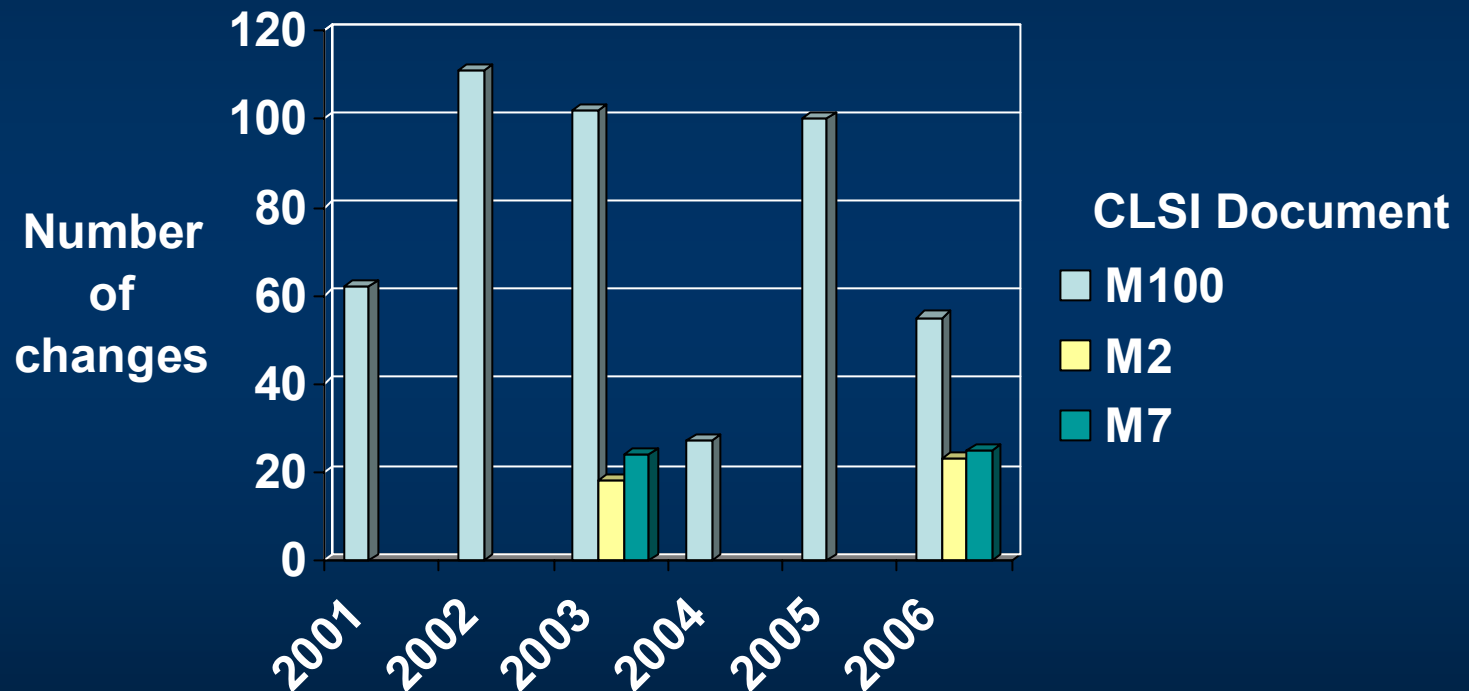
# ***Michigan Antibiogram QA Project: Part of the Initiative to Integrate Clinical Laboratories in Public Health Testing***

Martha Boehme, Project Lead  
Patricia Somsel  
Michigan Department of  
Community Health

# ***Objectives***

- **QUALITY IMPROVEMENT of AST**
  - Increase awareness/promote compliance with current standards of practice:
    - CLSI antimicrobial susceptibility testing standards
    - CLSI antibiogram guideline
- Develop/strengthen relationship with sentinel laboratories
- Develop effective training program for public health education of clinical laboratories

# *Changes in CLSI Recommendations 2001-2006*



Number and frequency of changes make it difficult for laboratory staff to keep current

# ***Michigan Clinical Laboratory Demographics 2002-2006***

- 110-115 sentinel labs
  - (fluctuates due to closures and mergers – we use 110 as “average” number)
- 53/110 (48%) in hospitals with < 100 beds
- Only 11 had PhD-level microbiologist on staff or available for consult



# ***Methods***

- Lab Survey 2003
  - 64/110 responses:
    - **>25% did not have newest (January 2003) CLSI document**
    - **8% of those did not even have 2002 document (M100-S12), which had many significant updates**
    - **One-third do not purchase CLSI documents yearly due to cost (~\$100)**
    - **41% are “not usually aware” that new documents have become available**

# ***Methods***

- Collect cumulative antibiograms from hospital laboratories
  - **Provides info about some of their testing practices**
  - **Provides info about their post-analytical practices**
  - **No additional burden for laboratory**
- Compare to CLSI recommendations
- Address gaps through educational offerings

# Antibiogram Example

Antimicrobials tested

Percent of organisms from specific group of patients over defined period of time that were susceptible to each antimicrobial agent

All bacterial isolates from sterile sites recovered from patients at Hospital XYZ from January 1, 2003 - December 31, 2004

## Gram-Negative

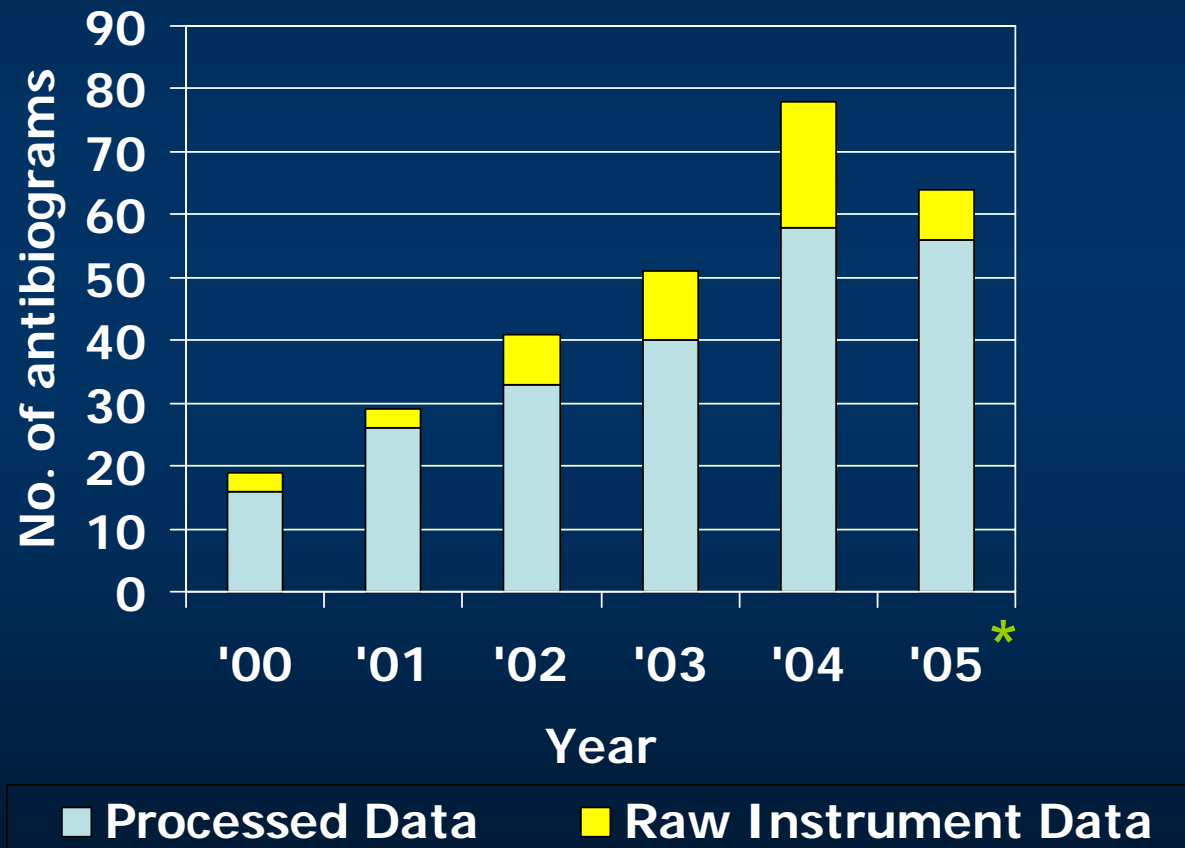
	# Isolates	Ampicillin	Ampicillin/sulbactam	Aztreonam	Cefazolin	Cefotaxime	Cefotetan	Ceftazidime	Ceftriaxone	Cefuroxime	Ciprofloxacin	Erythromycin	Gentamicin	Imipenem	Levofloxacin	Tetracycline	Ticarcillin	Tobramycin	Trimethoprim	Vancomycin
Escherichia coli	577	68	69	99	96	96	99	99	99	97	98		96	100	96	74	82	95	99	86
Enterobacter cloacae	32	13	34	82	3	74	58	78	78	50	100		100	100	100	78	92	74	100	100
Klebsiella pneumoniae	103	6	87	100	99	100	100	100	100	93	99		100	98	99	86	91	98	100	96
Proteus mirabilis	56	91	88	100	84	95	98	100	96	89	98		98	98	100	91	4	98	98	95
Pseudomonas aeruginosa	144			75			26	92	27		81		74	91	78	95		90	97	

## Gram-Positive

Staphylococcus aureus all	439			55							47	86			71	55				98	100
Staphylococcus aureus MRSA	198			0							14	76			17	0				97	100
Enterococcus faecalis	175	99									42	15	62*(s)		46						98
Enterococcus faecium	45	0									0	5	22*(s)		0						32
Streptococcus pneumoniae	42								99	80		71			94		68				100

Organisms

# *Number of Antibigrams Submitted Increased each year (2000-2005 data)*



\* Data still being submitted and analyzed

# ***Internal Antibiogram QI Assessment Tool***

- Developed uniform checklist based on CLSI M100 document
- Classified errors as major/minor
- Included only “processed” antibiograms (not raw instrument data summaries)
- Looked for errors, inclusion of select CLSI recommendations

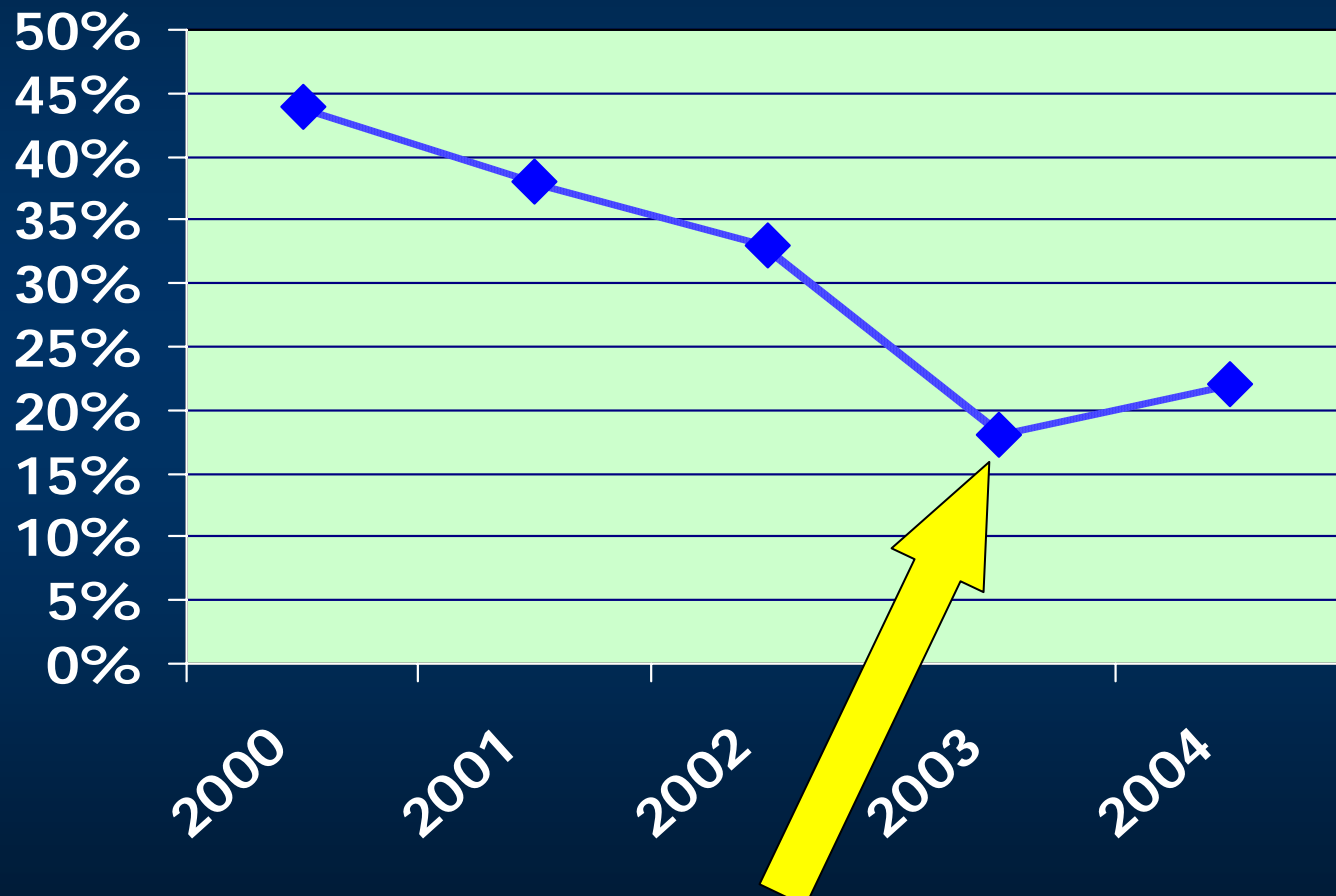
# Results

- **Examples of “major” errors:**
  - Unverified or unusual resistance patterns
    - e.g. *S. aureus* < 100% S to vancomycin
  - Inappropriate drugs reported
    - e.g. Oxacillin (instead of penicillin) reported on *Streptococcus pneumoniae*
  - Incongruent results
    - e.g. MRSA > 0% S to  $\beta$ -lactams
- **Examples of “minor” errors:**
  - Obvious math errors (e.g., 93% of 20 isolates!)
  - Organisms and/or /drugs misspelled

# *MDCH Actions to Address Gaps*

- Purchase and distribute CLSI AST-related documents to sentinel labs
- Provide educational offerings on AST topics:
  - MDCH workshops
  - MDCH lab newsletter and fax broadcasts
- Collaborate with SCACM at Michigan fall meetings and encourage participation by providing free registration for clinical labs

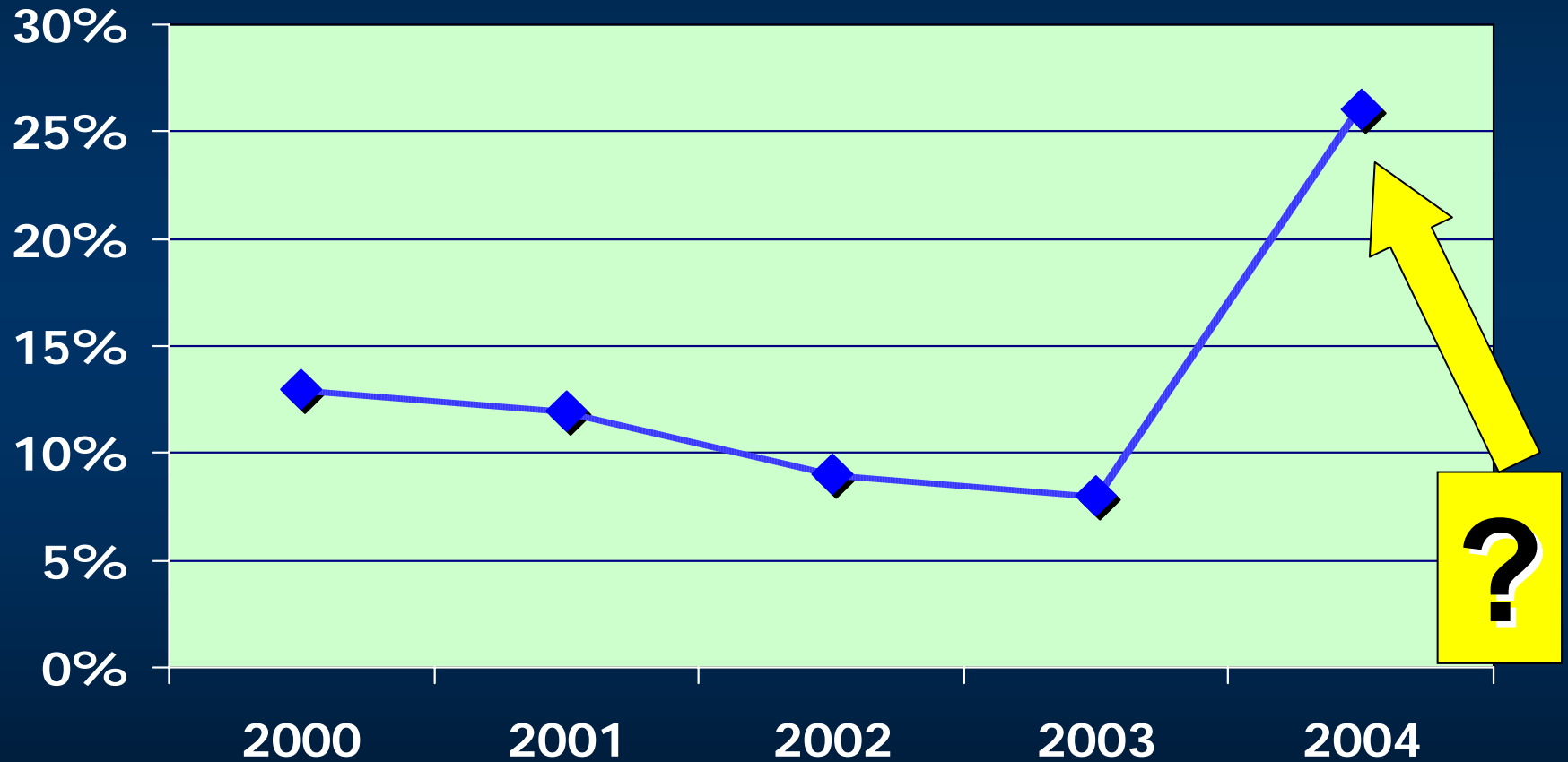
# *Percentage of AntibioGrams with Major Errors (2000-2004)*



**MDCH Education efforts in 2003 focused on AST issues**



# *Percentage of Antibio grams with Minor Errors (2000-2004)*



# **Many Improvements Noted over Life of the Project**

- Explanatory footnotes and comments increased
- *Streptococcus pneumoniae* data, dual (mening/ non-mening) interpretations were added
- ESBL data included
- Explanations of how data calculated, source of isolates
- Antimicrobial names spelled out or abbreviations defined

# *Conclusions*

- Clinical laboratories now rate “source for updates and documents” as one of most valuable services provided by MDCH
- Project established key “go-to” people at MDCH
  - 2004 Battelle study: 46% of labs made  $\geq 5$  inquiries of MDCH lab per year
  - Substantive inquiries to MDCH Integration Project coordinator increased 20-50% each year (2002-2006): 30-52% were AST-related

# ***Conclusions (cont'd)***

- Increase in antibiogram errors (2004 data) may be due to increased number of antibiogram submissions from new participants
- Ongoing outreach to clinical laboratories is essential to maintain progress
- Intangible benefits evident, though harder to measure:
  - Better working relationships, greater trust, ease in communications, more cooperation

# *Thank you*

- CDC - for supporting the Initiative to Integrate Clinical Laboratories in Public Health Testing
- Clinical Laboratory Staff in Michigan – for their dedication and commitment to the people they serve
- Exceptional MDCH Bureau of Laboratories staff - who recognize their responsibility to the citizens of MI for quality does not end at the walls of their laboratory.