



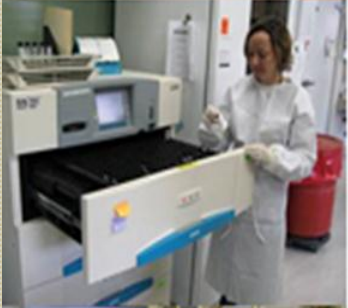
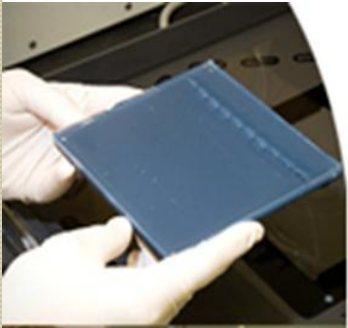
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# *How to Modify an FDA Approved Test without Really Trying-----and the Consequences*

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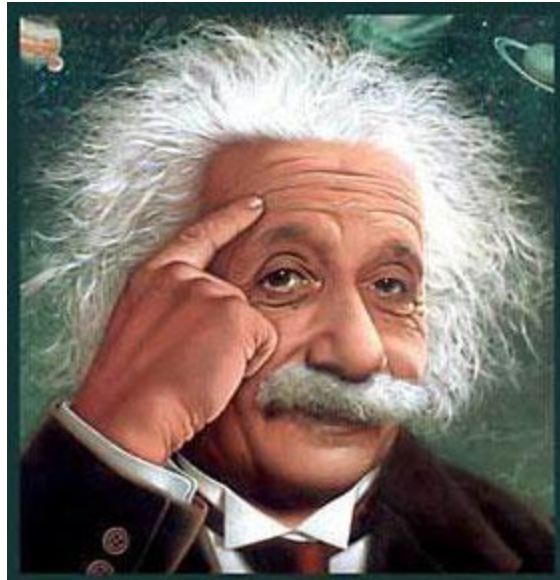
# My Laboratory's policy is

- A. that we use ONLY FDA approved/cleared molecular tests
- B. that we can use laboratory developed molecular tests



# Modification of an FDA approved test

“Modified by the laboratory” means any change to an assay that could affect its performance specifications





# Modification of an FDA approved test

- Using a specimen type not included in package insert
- Changing the extraction method
- Using a different collection device
- Changing specimen handling instructions
- Changing the cutoff value or method of calculation



# CLIA Requirements

After April 24, 2003, any new high complexity test introduced into the laboratory must be verified

- Laboratory developed test
- Modification of the manufacturer's test procedure
- Any non-FDA cleared method



# Terminology or

"What's in a name? That which we call a rose  
By any other name would smell as sweet."



- **Verification\***
  - The documentation of either commercial or laboratory developed tests to determine or confirm **test performance characteristics** before the test system is used for patient testing
    - A one-time process

\*Cumitech 31A

\*CLSI MM3-A2



# Terminology



## Validation

- The documentation that a test which has already been **verified** is repeatedly giving the expected results as the test is performed over a period of time.
  - **Quality control**
  - **Proficiency testing**
  - **Validation of employee competency**
  - **Instrument calibration**
  - **Correlation with clinical findings**





# Purposes of Method Verification

- To quantifiably characterize test performance
- To assess the potential for error
- To identify method-to-method differences
- To meet regulatory guidelines



# Verification Study Design for Laboratory Developed Tests (LDTs)





Does your laboratory have a policy and procedure for method verification?

A. Yes

B. No

C. If we do, I've never seen it



# CLIA Performance Characteristics

## Section 493.1253

CLIA specifies performance characteristics-----  
but not how to do it or criteria to be used

- Accuracy
- Precision
- Analytical sensitivity
- Analytical specificity
- Reportable Range
- Reference range(s)
- Any other characteristics required for test performance and interpretation of results



# Verification Study Design

- Establish the type and number of specimens necessary
- Decide on a comparative method or “gold standard”
- Acceptance criteria
- Methods for resolving discrepancies



# Discrepant Analysis

## Discrepancies

- May arise due to errors in the method being evaluated
- Comparative method is not 100% accurate

## Resolving discrepancies

- Use a designated reference standard method
- Send to another laboratory that uses a different method
- Use a test that targets another area of the gene



# What Types of Samples can be used?

Should be typical of those that will be routinely tested

- Patient samples with known results
  - Retention specimens
  - Specimens from another laboratory

## Other Options

- Quality control material
- Proficiency testing samples
- Calibration material
- Spiked negative patient specimens
- Manufacturer's verification panels???



# PERFORMANCE CHARACTERISTICS FOR VERIFICATION







# Analytical Accuracy

Closeness of an individual measurement to the "true" value, as determined by a reference method

Numbers of known positive and negative specimens should be balanced or statistically significant to have confidence in the test result

- e.g.  $\geq 50$  positive,  $\geq 50$  negative
- Confidence interval of 78-97% (CLSI EP12)

$$\frac{\text{Number of correct results}}{\text{Total number of results}} \times 100$$



# Side-by-Side Comparison of Real-Time PCR Assay to Gen Probe® Amplified™ MTD Test at WSLH

<b>Number of Specimens</b>	<b>MTD Positive</b>	<b>MTD Negative</b>	<b>MTD Inhibited</b>
<b>Real-Time PCR Positive</b>	<b>49</b>	<b>1</b>	<b>2</b>
<b>Real-Time PCR Inhibited</b>	<b>0</b>	<b>0</b>	<b>0</b>
<b>Real-Time PCR Negative</b>	<b>0</b>	<b>50</b>	<b>3</b>



$$\text{Accuracy} = (99/100) \times 100 = 99\%$$

1 “Incorrect” Result

### Discrepant Analysis

- A different TB NAAT
- Culture Result



# Analytical Sensitivity (Limits of Detection)

**LoD---the lowest amount of target that can be detected by a test system with a stated probability**

## Methods

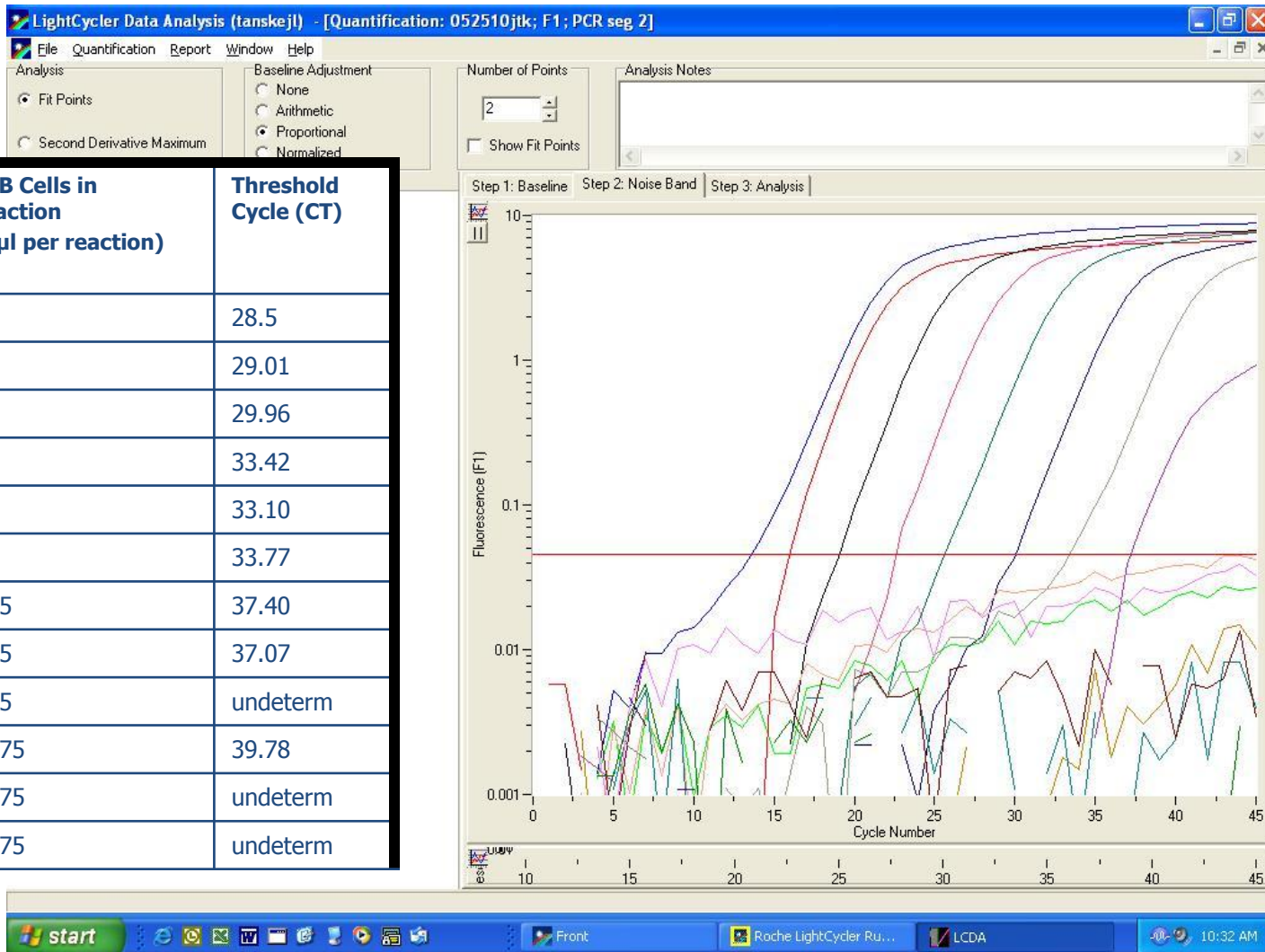
- Spiking whole organisms into negative specimens
  - Known CFUs/ml, PFUs/ml, TCID<sub>50</sub>/ml
- Spiking with known number of copies of the target

## Evaluate for the influence of microbial diversity

- Different patient isolates, serogroups, serotypes, lineages, resistance phenotypes, etc.

Reference---EP17-A: Protocols for Determination of Limits of Detection and Quantitation

# Anaytical Sensitivity





Cell Suspension	Mtb/Reaction	Threshold Cycle
1.5 X 10 <sup>3</sup>	7.5	32.98
	7.5	33.57
	7.5	33.81
	7.5	32.89
	7.5	32.96
	7.5	31.9
	7.5	33.2
	7.5	30.9
	7.5	33.00
	7.5	31.45
	7.5	33.57
	7.5	32.48
	7.5	31.75
	7.5	30.12
	7.5	32.58
	7.5	33.42
	7.5	31.69
	7.5	33.01
	7.5	32.79
7.5	32.51	
<b>Average</b>		<b>33.11</b>

1.5 X 10 <sup>2</sup>	0.75	36.47
	0.75	36.12
	0.75	35.70
	0.75	35.70
	0.75	34.38
	0.75	35.77
	0.75	38.38
	0.75	35.88
	0.75	36.86
	0.75	37.45
	0.75	35.22
	0.75	36.13
	0.75	37.13
	0.75	35.56
	0.75	37.62
	0.75	38.10
	0.75	37.00
	0.75	36.51
	0.75	Undeterm
0.75	36.08	
<b>Average</b>		<b>36.42</b>

LoD



# Analytical Specificity

**Ability of a method to detect only the intended target**

Verify for cross-reactivity

- Organisms closely related to the target organism
- Organisms that represent normal flora of the specimen being tested
- Organisms that cause similar disease syndromes

Can use whole organisms that go thru the complete method and/or extracted nucleic acids

# Specificity and Breadth of Detection



## Negative PCR Result

*M. avium* complex  
ATCC700898

*M. goodnae* ATCC 1218

*M. xenopi*

*M. scrofulaceum*

*M. peregrinum* ATCC 700686

*M. smegmatis* ATCC 1546

*M. marinum*

*M. mucogenicum*

*M. chelonae*

*M. abscessus*

*M. fortuitum* ATCC 1447

*M. kansasii* ATCC 12478

*Tsukamurella* sp.

*L. pneumophila*

*N. meningitidis*

*S. pneumoniae*

Group A *Streptococcus*

*H. influenzae*

*P. aeruginosa*

*K. pneumoniae*

*B. Pertussis*

*B. parapertussis*

## Positive PCR Result

MTBC ATCC 27294

MTBC ATCC 35828

32 TB patient isolates and  
3 *M. bovis* BCG isolates





# Interfering Substances

- Blood

- Lipemic
- Icteric
- Hemolyzed
- Anticoagulants (EDTA, heparin, ACD)

- Sputum

- Bloody
- Decontamination process



# Precision (Reproducibility)

Within run

Run-to-run, same day

Day-to-day, different analysts

Inter-instrument

Protocol described in CLS EP12

- e.g. For a qualitative assay interpreted from a quantitatively measured signal-----Calculate SD and CV from 10-20 day-to-day quality control results
- Test 2 concentrations in duplicate, 2x/day for 20 days
- Run aliquots of a single specimen 30 times in a single run
- Panel of samples run by different analysts

Goal of 95% typical for PCR assays



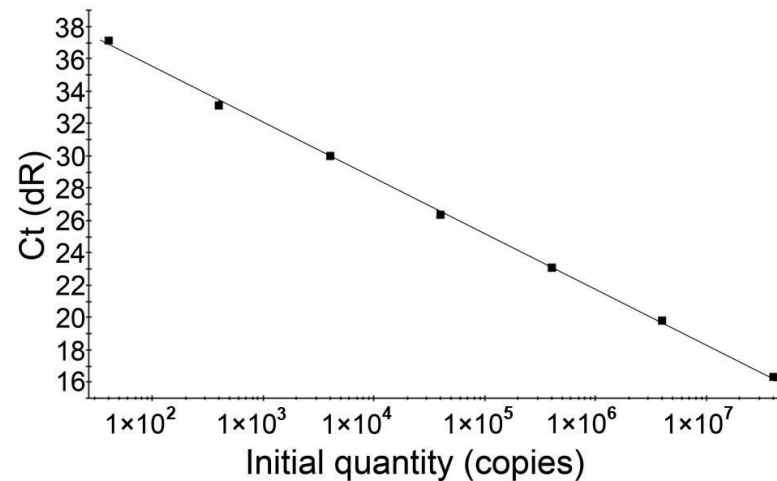
# Reportable Range

For qualitative assay---Not Applicable

- Detected or not detected

For quantitative assay

- The range of results for which a test has been proven to yield numerically accurate results. (CLSI Document EP17-A)





# Verification Documentation

## Write up

- Purpose and Background
  - References for clinical utility
  - Intended use of the assay
  - Specimen types and matrixes
- Methods
- Results
- Conclusions

Review, approval, sign off by Laboratory Director or designee who qualifies as a Director

Save for  $\geq 2$  years after test is discontinued



# FDA-Cleared Test Verification

If unmodified

- Accuracy
- Precision
- Reportable range
- Reference range

**Don't have to assess sensitivity and specificity**

Accuracy and precision should fall within manufacturer's specifications

- Can use fewer specimens



# Public Health Laboratory Exceptions

Exceptions for PHLs when calibration or control materials are not available

- e.g. During public health emergencies, tests for emergent diseases, or public health threats
- Emergency Use Authorization (EUA)



# Public Health Laboratory Exceptions con't

## Given temporary CLIA exemption

- Must follow CDC protocols w/o modification
- Personnel must show proficiency in the test method
- Must document alternative methods to show accuracy
  - Send a number of samples to CDC for verification
  - Test with another method

Must do complete validation when calibration material is available or EUA expires



# Summary

Verification studies are a “Necessity of Laboratory Life”

Verification studies require a major time commitment

Verification studies can be a major expense

Verification studies must be carefully designed

- Important to utilize the guidelines that are available





# References

CLSI document MM3-A2. Molecular Diagnostic Methods for Infectious Diseases.

CLSI document MM6-A. Quantitative Molecular Methods for Infectious Diseases.

CLSI document MM13-A. Collection, Transport, Preparation, and Storage of Specimens for Molecular Methods.

CLSI document MM17-A. Verification and Validation of Multiplex Nucleic Acid Assays.

CLSI document EP12-A2. User Protocol for Evaluation of Qualitative Test Performance.



# References

CLSI document EP17-A. Protocols for Determination of Limits of Detection.

CLSI EP05-A2 document. Evaluation of Precision Performance of Quantitative Measurement Methods.

CUMITECH 31. Verification and Validation of Procedures in the Clinical Microbiology Laboratory. ASM Press

National Laboratory Training Network Verification of Infectious Disease Molecular Assays. August 2005.

Copies available [www.nltn.org](http://www.nltn.org)



# References

CLSI document GP27-A2. Using Proficiency Testing to Improve the Clinical Laboratory.

CLSI document GP29-A. Assessment of Laboratory Tests When Proficiency Testing is Not Available.



Thank you!