## **STEC Whole Genome Sequencing Project**

#### Eija Trees, Ph.D., D.V.M.

#### Chief, PulseNet Next Generation Subtyping Methods Unit

16<sup>th</sup> Annual PulseNet Update Meeting August 29<sup>th</sup>, 2012



National Center for Emerging and Zoonotic Infectious Diseases Division of Foodborne, Waterborne and Environmental Diseases

## **Objectives of the Presentation**

- Describe the STEC genome sequencing project
- Describe the sequence diversity seen in the STEC population
- Way forward

## Outline

- Background and Objectives of the Project
- Materials and Methods
- Results
- Conclusions
- Future Plans
- 100K Pathogen Genome Sequencing Project

## Background

## **CDC Bioinformatics Blue Ribbon Panel in June 2011**

### **2011 end-of-year funding from CDC OD, OID and NC**

- Implemented a proof-of-concept bioinformatics core group through an outside contract
  - In 2012 focus on two pilot projects:
    - MicrobeNet: development of shared bioinformatic services and web platforms
    - Next Generation PulseNet: Whole genome sequencing, comparative genomics, molecular epi

## Next Generation PulseNet Project Objectives

- Perform whole genome sequencing, assembly and comparative analysis of 250 STEC strains
  - Reference database
- Understand diversity in the STEC population and determine correlation between WGS data, PFGE and epi
- Mine the resulting data for targets to predict strain type, virulence and antimicrobial resistance quickly and at high resolution

## Next Generation PulseNet Project Objectives (cont'd)

## Public health benefits:

- Standardized workflows/SOPs for next generation sequencing across multiple platforms
- Reusable tools, techniques and software "pipelines" for a wide range of comparative genomics and molecular epidemiologic applications
- Builds capacity for rapid genome-level molecular epidemiology

## **MATERIALS AND METHODS**

## **Strain Selection**

# Selection criteria should help us to understand variation:

- 1. within an outbreak or during a carrier state
- 2. among epidemiologically unrelated isolates within a serotype and between serotypes

## Strain selection (cont'd)

#### Variation within an STEC outbreak or a carrier state

- 10 isolates each from 5 outbreaks
  - O157:H7 (3), O111:NM (1), O145:NM (1)
  - Different types of outbreaks: clonal vs. polyclonal, short vs. long lasting, point source vs. vehicle never identified
- 11 isolates recovered from the same patient in the course of 2<sup>1</sup>/<sub>2</sub> months

## Strain selection (cont'd)

# Variation among epidemiologically unrelated STEC strains

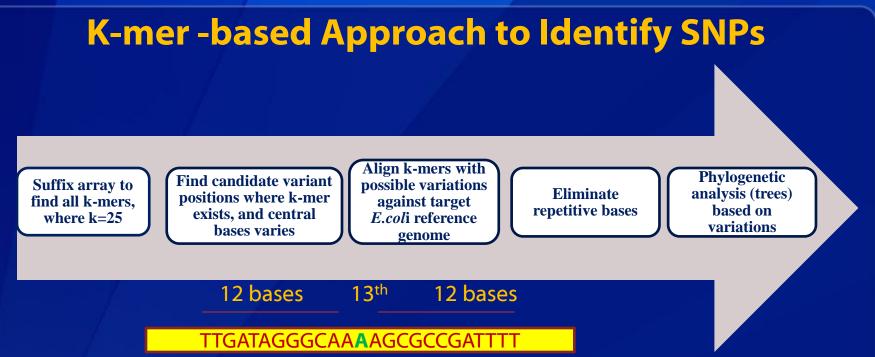
- 25 strains each from serogroups O26, O111, and O121
- 5 strains each from serogroups O45, O69, O91, O103, O118, O145
- 1 strain each from serogroups ranked in prevalence #11-22
- 1 representative each from past historical O157 (and non-O157) outbreaks (36 in total)
- Sporadic O157 isolates representing common PFGE patterns (23 in total)

Other E. coli pathotypes: ETEC (6), EPEC (3), EAggEC (3), EIEC (1)

## Whole genome sequencing and assembly

#### Illumina HiSeq 2x100 bp; average coverage 400x

- Pros: highly accurate reads with massive coverage
- Cons: reads short
- Assembled with CLC Bio; average of 200 contigs
- Subset of 99 isolates sequenced with Pacific Biosciences SMRT (on-going)
  - Pros: longer reads (up to 10 kb)
  - Cons: error rate high, multiple SMRT cells required per strain for adequate coverage
  - Hybrid assembly with error correction using Illumina reads being optimized



- When k-mer 25 is used, the 13<sup>th</sup> base position is used for determining if there is a variation
- No prior sequence assembly or multiple sequence alignment required
- Fast completion of analysis
- Phylogenies less affected by regions that have undergone strong selection, deletions or horizontal gene transfer
- Dependent on high quality data
- Reference genome required

## **RESULTS**

## Probing Diversity within an Outbreak – 0157:H7 in MN Daycare (2008)

PFGE-Xbal	PFGE-BInI	_
		2011EL-2091 2011EL-2098
Xbal/BlnIPFGE VARIANT		2011EL-2101
MLVA VARIANT		2011EL-2094 2011EL-2090
		2011EL-2092 2011EL-2093
Bini PFGE VARIANT		2011EL-2097 2011EL-2099
	1 5 5 1 101 0 0 0 0 0	2011EL-2096

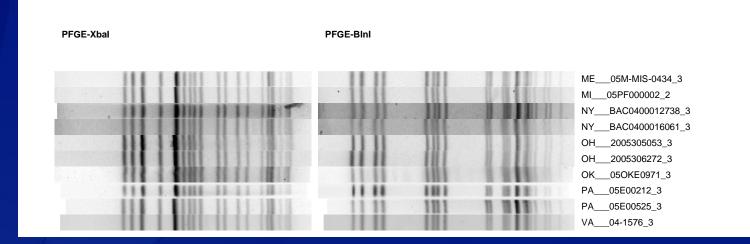
Relatively clonal short lasting point source outbreak

#### Probing Diversity within a Clonal Point Source O157 Outbreak – MN Daycare

Color Key								0.0	0.5		1.0	1.5
AT G C								Reference -	Xba-Bln variar	nt	·	
			_				2083 1545 1944 1944 2039 1955 2039 2039 2039 2045	X2011EL.2091 —				
							21853 21859 21859 21859 21854 21854 21853 21854 21855 21855 21855	X2011EL.2092 -				
							933 17345 492 2647 2462 7462 71427	X2011EL.2093 —				
							01211 01229 01285 01285 01265 01242 22424 22424 01240	X2011EL.2098 —				
							368 1933 2930 1944 1943 1943 1945	X2011EL.2096 –	MLVA variant			
		E 4			2 0	g	1929 1920 1925 2949 2949 2949	X2011EL.2094 -	Bln-variant			
	Referenc	1EL.2101	1EL.2094	1EL.2099	1EL.209 1EL.209	1EL.209	1EL 209	X2011EL.2099 -				

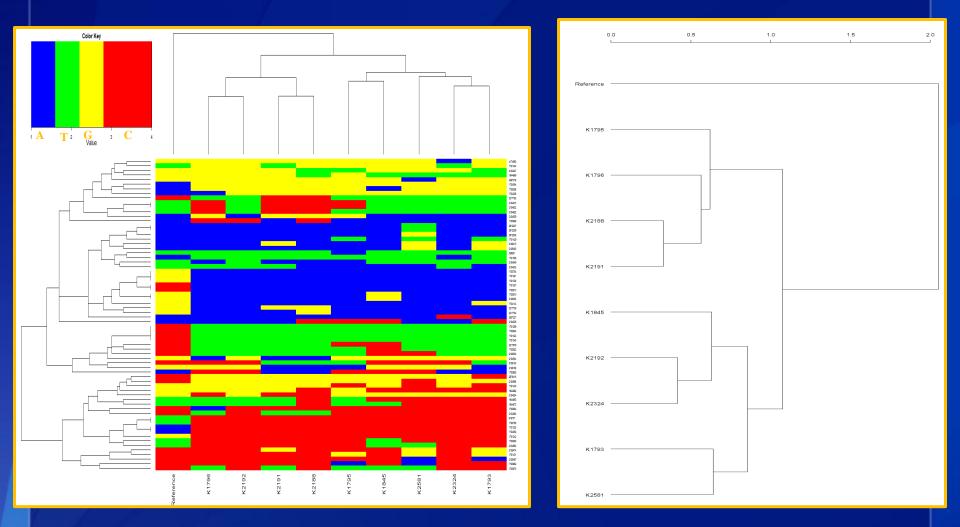
**Reference Sample:** 2011EL-2090\_O157:H7\_EXHX01.0589\_EXHA26.1376\_main MLVA • The total number of SNPs found across all the samples from this outbreak at 100% frequency is **39**.

## Probing Diversity within Clonal Long Lasting 0157:H7 Outbreak (2004-2005)



- 110 2-enzyme matches in 20 states within 15 months
- Rare PFGE type
- An indistinguishable new MLVA pattern
- Vehicle never identified

#### Probing Diversity within a Clonal Long Lasting O157 Outbreak – Not resolved



Reference Sample: K1792\_O157:H7\_EXHX01.0086\_EXHA26.0576

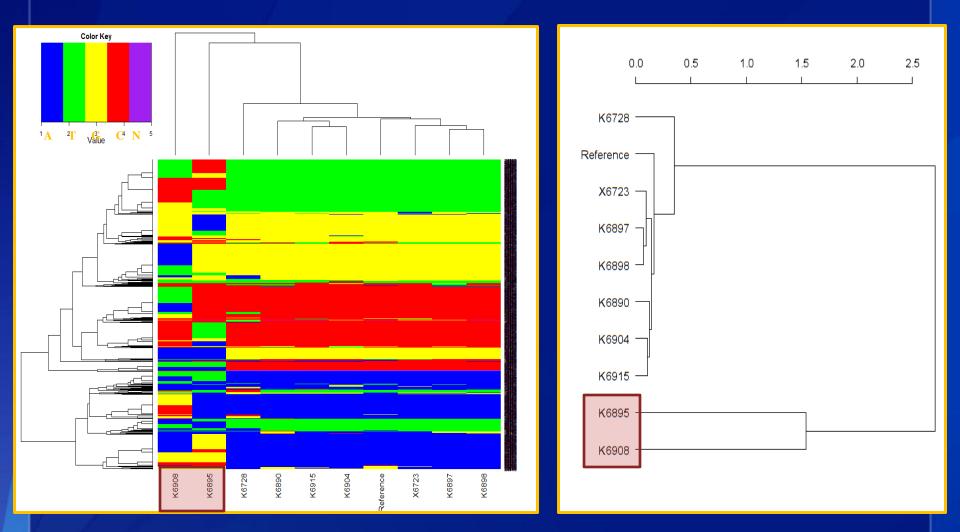
• The total number of SNPs found across all the samples from this outbreak at 100% frequency is 68

## Probing Diversity within Polyclonal Outbreak – O111:NM in OK Restaurant (2008)

PFGE-Binl+PFGE-Xbal Xbal & Binl	PFGE-Xbal	PFGE-Bini				
100 -90 -90			Key	Serotype	PFGE-Xbal-pattern	PFGE-BInI-pattern
			OK08OKE1082A	E. coli O111:NM	EXDX01.0050	EXDA26.0174
			OK08OKE1248	E. coli O111:NM	EXDX01.0050	EXDA26.0174
			OK08OKE1296	E. coli O111:NM	EXDX01.0050	EXDA26.0174
			OK08OKE1310	E. coli O111:NM	EXDX01.0050	EXDA26.0174
			OK08OKE1314	E. coli O111:NM	EXDX01.0050	EXDA26.0174
			OK08OKE1079B	E. coli O111:NM	EXDX01.0175	EXDA26.0174
			OK08OKE1244	E. coli O111:NM	EXDX01.0337	EXDA26.0190
			OK08OKE1115	E. coli O111:NM	EXDX01.0327	EXDA26.0183
			OK08OKE1265	E. coli O111:NM	EXDX01.0320	EXDA26.0177
			OK08OKE1308	E. coli O111:NM	EXDX01.0334	EXDA26.0043

- 1 Xba variant, 4 Xba-Bln variants
- Vehicle not identified

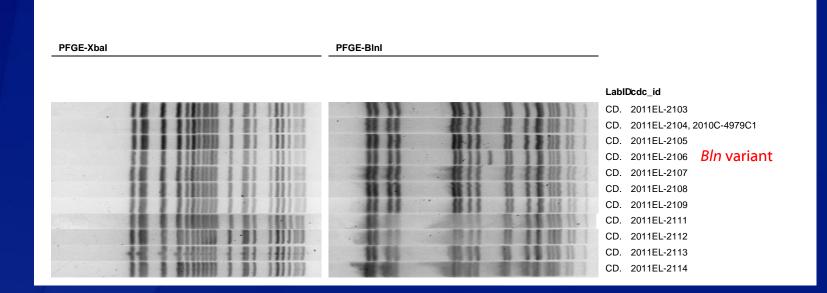
#### **Probing Diversity within Polyclonal O111 Outbreak – OK Restaurant**



**<u>Reference Sample: K6722\_O111:IsolatetoCDC\_EXDX01.0050\_EXDA26.0174</u></u>** 

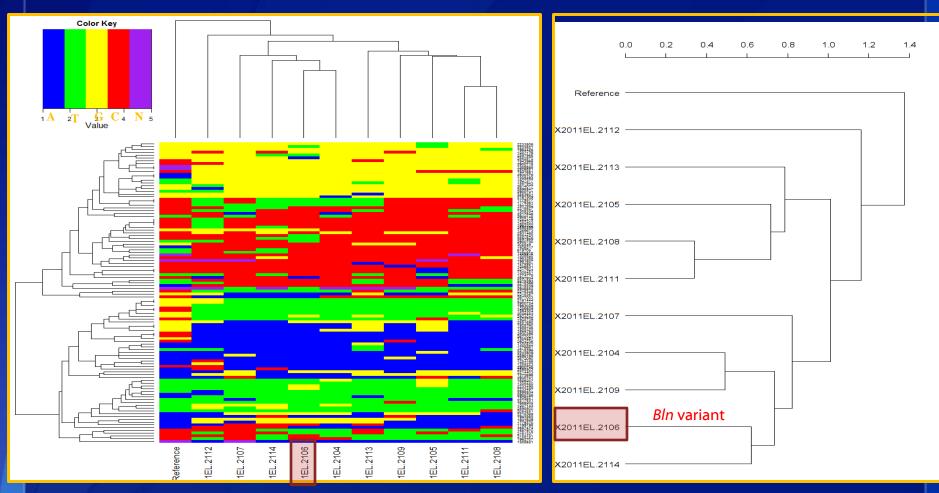
• The total number of SNPs found across all the samples from this outbreak at 100% frequency is 947

## **Probing Diversity During a Carrier State**



- Collected from the same person within a period of 2<sup>1</sup>/<sub>2</sub> months
- An indistinguishable MLVA pattern

#### Probing Diversity during a Carrier State – Serial Isolates from the Same Person

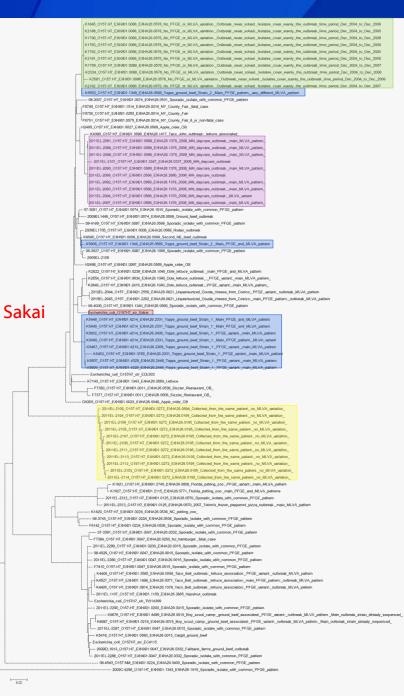


**<u>Reference Sample: 2011EL-2103\_O157:H7\_EXHX01.0272\_EXHA26.0195</u>** 

• The total number of SNPs found across all the samples from this "outbreak "at 100% frequency is 108

## K-mer –based **Phylogenetic Tree for STEC** 0157

93 to 2101 **SNPs** compared to the reference



**Outbreak 1** (Not solved, 2004-05) Outbreak 2 (Daycare, 2008) Outbreak 3

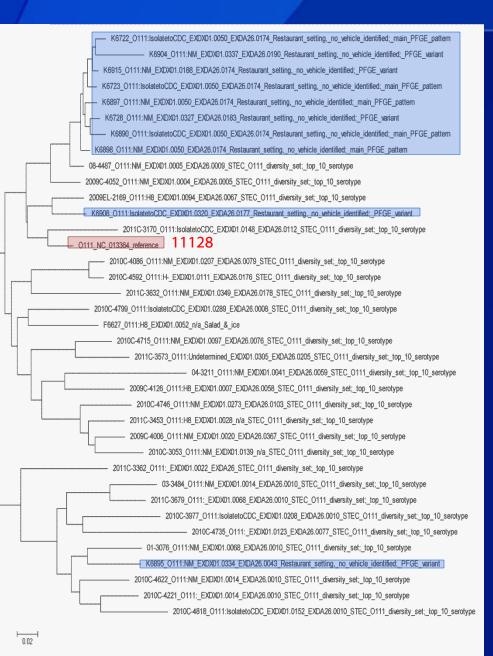
(Topps ground beef, 2007)

Long term carrier patient

0.02

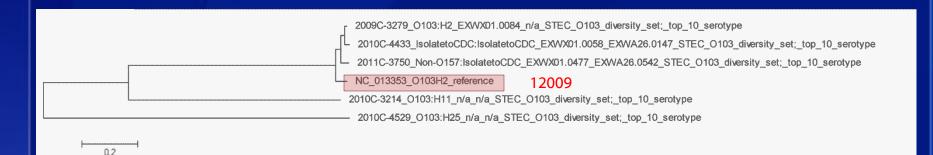
## K-mer -based Phylogenetic Tree for STEC 0111

195 to 1229 SNPs compared to reference



#### Restaurant outbreak, 2008

## K-mer -based Phylogenetic Tree for STEC 0103



1789 to 20,094 SNPs compared to the reference

## **Conclusions Based on the Illumina Data**

- K-mer –based clustering appears to have good correlation with epidemiological and PFGE data
- Variation within a (relatively) clonal outbreak < 100 SNPs
- The variation between unrelated strains of the same serotype hundreds to ~ 2000 SNPs
- The variation between serotypes > 10,000 SNPs

## **Future Plans**

#### Improve the assemblies using PacBio sequences

- Make the draft genomes publicly available
- Additional sequencing to close reference genomes
- Define a cross serotype gene set to be used to determine strain subtype, virulence and antimicrobial resistance profiles ('super' MLST)
  - Presence / absence
  - SNPs

Add prospective strains from outbreaks to the database

## **Questions to be Answered**

### Which clustering method will work for surveillance?

- 1. K-mer based clustering as a primary tool with gene-based 'super' MLST as a secondary tool OR
- 2. Gene-based 'super' MLST as a primary tool

#### How to standardize data across the platforms?

Different error profiles

How to get an actionable report out from raw reads?

## **100K Pathogen Genome Sequencing Project**

- Dilemma: the safety and security of the world food supply is hindered by lack of food-related genomes
- Partnership between BGI and UC Davis
  - <u>http://100kgenome.vetmed.ucdavis.edu</u>
- Goal: sequence 100,000 foodborne pathogen isolates within 5 years
  - Sequences will be made publicly available

## Steering committee

- BGI
- UC Davis School of Veterinary Medicine
- FDA
- CDC
- Agilent Technologies, Inc.
- Mars, Inc.

## 100K Pathogen Genome Sequencing Project (cont'd)

#### **Isolate priority list**

🖉 🛛 Tier 1	Tier 2	Tier 3
Salmonella	Yersinia	Toxigenic bacilli
Campylobacter	Shigella	Norovirus
E. coli	Clostridium	Hepatitis
Vibrio Listeria	Enterococcus Cronobacter	Rotovirus

• Seeking isolates from diverse types of food, distinct environments, diverse regions of the world and longitudinal isolate collections

## Acknowledgements

- CDC/EDLB
  - Peter Gerner-Schmidt
  - John Besser
  - Efrain Ribot
  - Nancy Strockbine
  - Cheryl Tarr
  - Lee Katz
  - Amber Schmidtke
  - Ashley Sabol
  - Haley Martin
  - Devon Stripling
- MN Dept. of Health
  - Dave Boxrud

- CDC Bioinformatics Core Group
  - Duncan MacCannell
  - Ryan Weil
  - Shankar Changayil
  - Satishkumar Ranganathan
  - Kun Zhao
- Expression Analysis
  - Stephen Sifed
  - Victor Weigman
  - Steve Mcphail

## Thank You for Your Attention! Questions?

For more information please contact Centers for Disease Control and Prevention

1600 Clifton Road NE, Atlanta, GA 30333 Telephone: 1-800-CDC-INFO (232-4636)/TTY: 1-888-232-6348 E-mail: cdcinfo@cdc.gov Web: http://www.cdc.gov

The findings and conclusions in this report are those of the authors and do not necessarily represent the official position of the Centers for Disease Control and Prevention.



National Center for Emerging and Zoonotic Infectious Diseases Division of Foodborne, Waterborne and Environmental Diseases