



One test. Answers for Life.

Second-tier DNA Confirmation of Newborn Screening by Targeted Next Generation Sequencing

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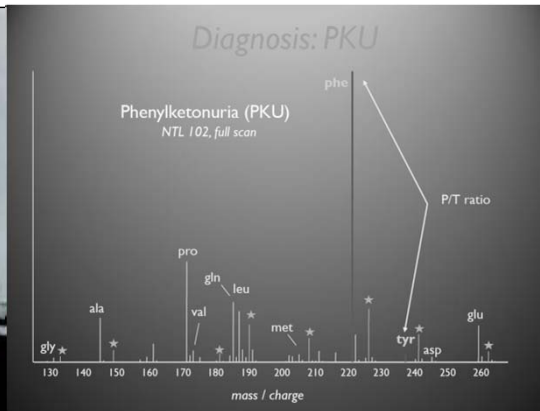
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Newborn Screening & Clinical Genomics

1961

1990's

2010-2012



Robert Guthrie develops simple Newborn Screening (NBS)

Development of automated MS/MS screening across several disorders

Current *de facto* standard

SCIENCE VOL 330 17 DECEMBER 2010

BREAKTHROUGH OF THE YEAR | NEWSFOCUS

HUMAN GENOMICS

Carrier Testing for Severe Childhood Recessive Diseases by Next-Generation Sequencing

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OPEN ACCESS Freely available online

PLoS one

Genetic Mapping and Exome Sequencing Identify Variants Associated with Five Novel Diseases

RESEARCH ARTICLE

DIAGNOSTICS

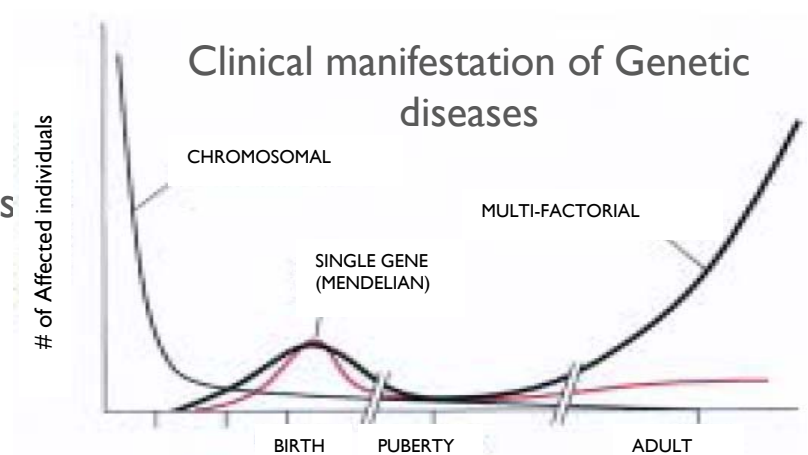
Rapid Whole-Genome Sequencing for Genetic Disease Diagnosis in Neonatal Intensive Care Units

Carol Jean Saunders,^{1,2,3,4,5*} Neil Andrew Miller,^{1,2,4*} Sarah Elizabeth Soden,^{1,2,4*} Darrell Lee Dinwiddie,^{1,2,3,4,5*} Aaron Noll,¹ Noor Abu Alnadi,⁴ Nevene Andrews,³ Melanie LeAnn Patterson,^{1,3} Lisa Ann Krivohlavek,^{1,3} Joel Fellis,⁶ Sean Humphray,⁶ Peter Saffrey,⁶ Zoya Kingsbury,⁶ Jacqueline Claire Weir,⁶ Jason Betley,⁶ Russell James Grocock,⁶ Elliott Harrison Margulies,⁶ Emily Gwendolyn Farrow,¹ Michael Artman,^{2,4} Nicole Pauline Safina,^{1,4} Joshua Erin Petrikin,^{2,3} Kevin Peter Hall,⁶ Stephen Francis Kingsmore^{1,2,3,4,5†}

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Why Newborn Genomics?

- **Mendelian Diseases disproportionately affect Newborns**
 - ~3500 genetic diseases with molecular basis
 - >10% of NICU admissions are genetic
 - Current NBS tests limited to 29+ diseases
 - 2nd tier DNA testing to validate biochemical results
- **Advantage of NGS based DNA testing**
 - Find causal variants (rare/novel) in gene(s)
 - A 'universal' NGS approach avoids repeated, serial single gene testing
 - Current Sanger sequencing is expensive (\$3-10K) and slow (3 months to 1 year)



Gelehrter TD, Collins FS, Ginsburg D. Principles of Medical Genetics. 2nd ed. Baltimore, MD: Williams & Wilkins; 1998:1-42

NICU- Neonatal Intensive Care Unit
NBS-Newborn Screening
NGS-Next Generation Sequencing

Why Targeted (Exome) Sequencing for now?



Test Menu	Cost (\$)	Throughput	Fold Efficiency
Whole Genome (Res.)	7,666*	1	1
Exome (Res)	1,200	15	95
Neonate Panel (Clinical)	<1000	150	>1140

- Majority of known disease-causing mutations in exons
- Exome = protein-encoding parts of genes
- **Targeted NGS is Cost & Throughput Efficient**

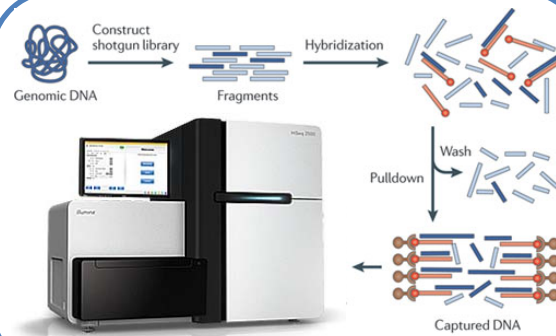
*Saunders et al., (2012) Rapid Whole Genome Sequencing for Genetic Disease Diagnosis in NICUs

Workflow for 2nd Tier Newborn Screening

Sample Isolation **2h**



DNA Capture & Sequencing **92h**



Raw Data Management **10h**

```
AGGTCGTTACGTTACGCTAC
GACCTACATCAGTACATAG
GCATGACAAAGCTAGGTGT
```

Analysis & Interpretation **1h+**

Omicia Opal - Beta 0.10.0

VAAST Trio Report

Overview
 Proband: Daughter
 Unaffected Mother: Mother
 Unaffected Father: Father
 Background: UK Project Phase 1
 VAAST Release: RCT 2 (recessive model)

Var	Ref	Change	Proband	Father	Mother	Effect	Global	Omicia	V-Score	O-Score	Evidence
Unknown	38524	chr7 385815 G1138822	c466_565delT g.AacG2h	hom	het	het	beneschr	beneschr	0.408	30.74	30.74 al
Unknown	38525	chr7 3770570 G1138823	c105_205delA g.AcG3h	hom	het	het	beneschr	beneschr	0.262	30	30 al
Unknown	38526	chr7 43945 G1138824	c121_121delC p.Leu17h	hom	het	het	beneschr	beneschr	0.133	29.32	29.32 al
Unknown	38527	chr7 866514	c1135_1135del g.TG17h	hom	het	het	nonbeneschr	beneschr	0.362	27.67	27.67 al
Proband	38528	chr7 2101110	c480G-C g.GG18h	het	-	het	non-syn	-	0.87	12.95	27.24 al
Proband	38529	chr7 7205942	c454G-A g.GG17h	het	-	het	non-syn	-	0.88	10.29	27.24 al
Unknown	38530	chr2 4009259	c122C-T p.Trp18h	hom	het	het	non-syn	-	0.376	24.36	24.36 al

8 samples, 105 Hrs, <\$10,000 = Real Neonatal Genomics!

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Workflow for 2nd Tier Newborn Screening

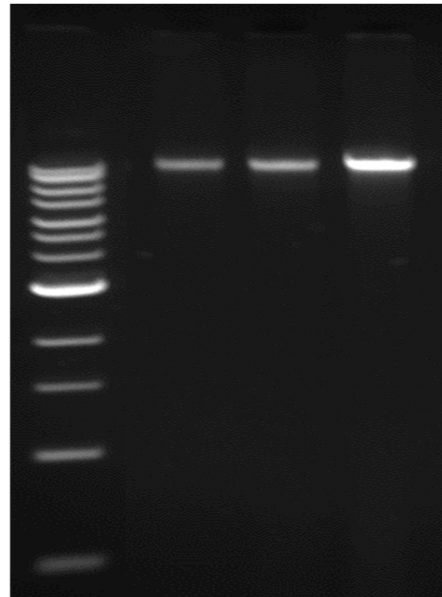
Sample Isolation

2h

- High M.Wt. DNA
- PCR Amplifiable and NGS ready
- Processing Time ~ 2 hrs
- Amenable to automation

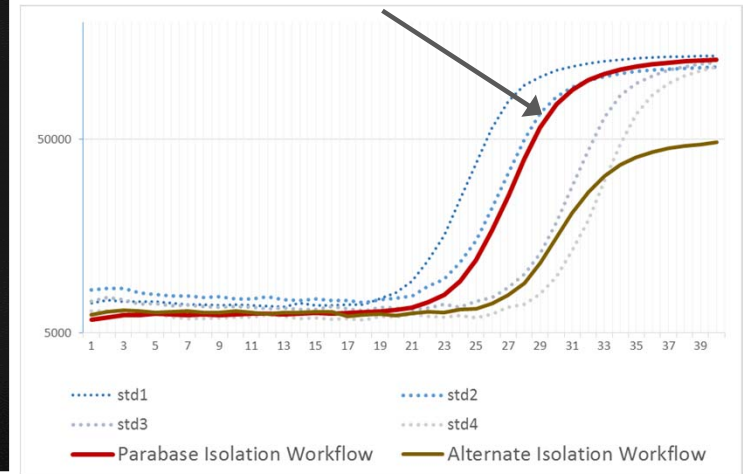


%DBS 25 50 100



Yield 400ng per Spot

More yield and DNA purity than alternate workflow



Workflow for 2nd Tier Newborn Screening

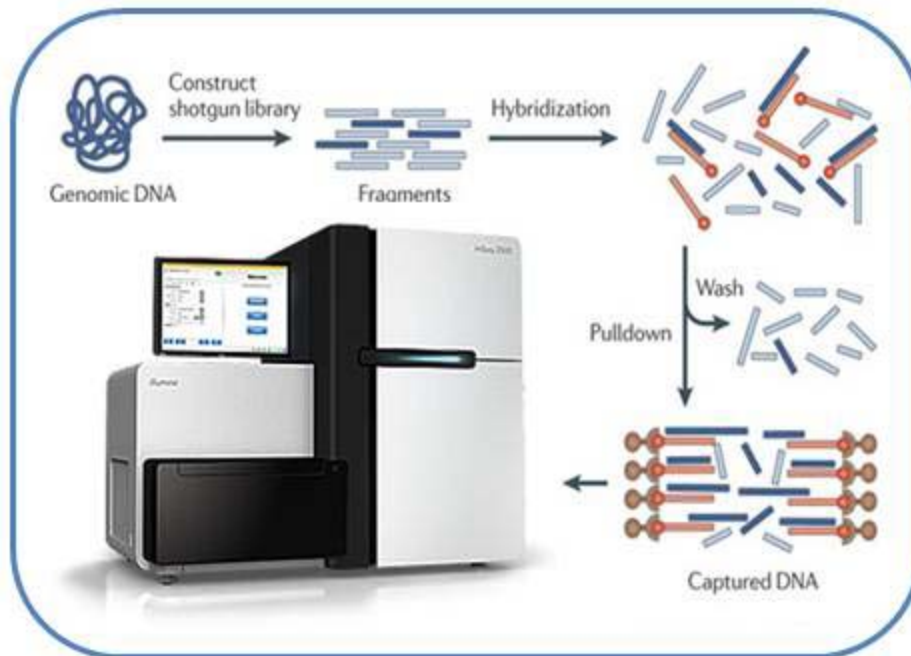
DNA Capture* & Sequencing**

92h

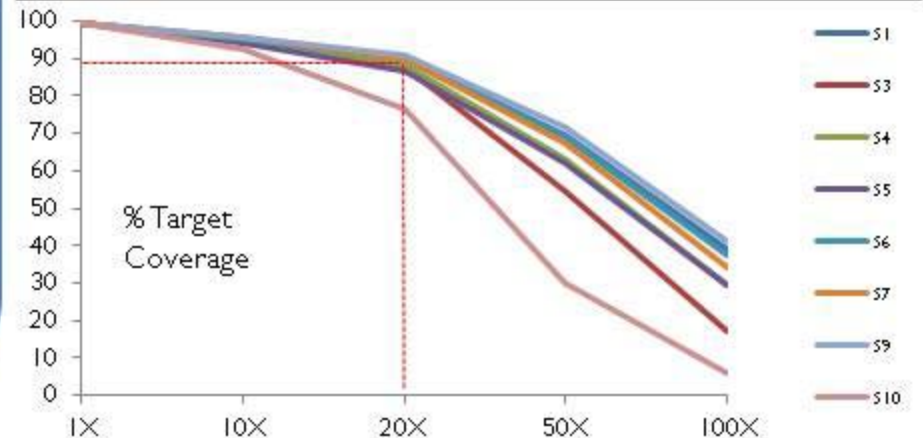
* Room for improvement

** Process reduced from 19* days to 92 hrs

- Library Prep ~ 5 hrs
- Capture ~ 66 hrs
- HiSeq 2500 PE 75 ~ 21 hrs
- Fast turnaround & High quality data



	S1	S3	S4	S5	S6	S7	S9	S10
TOTAL READS (Mln)	92	68	77	75	90	85	96	49
MAPPED READS (Mln)	89	58	72	72	86	79	91	37
% READS MAPPED	96	86	95	96	95	95	95	76
READS ON TARGET (Mln)	69	38	56	56	66	61	71	21
% READS ON TARGET	74	56	73	75	73	73	74	42



Data generated on Nimblegen Exome and HiSeq2500

Variant calling GATK2 (Broad Institute, MA) by CFI and Real Time Genomics

2 Known (MSUD and PA) & 8 blinded samples were provided by Clinic for Special Children, PA (S1-S10)

*Saunders et al., (2012) Rapid Whole Genome Sequencing for Genetic Disease Diagnosis in NICUs

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Workflow for 2nd Tier Newborn Screening

Raw Data Management
(Mapping, alignment & variant calling)

10h

```
AGGTCGTTACGTACGCTAC
GACCTACATCAGTACATAG
GCATGACAAAGCTAGGTGT
```

FASTQ



.vcf (PG/CFI or RTG)



Omicia

- We have integrated and tested two fast data management and processing workflows
 - Clinical Future Inc. (CFI)-uses GATK2
 - Real Time Genomics (RTG)- proprietary
- All Variant (.vcf) files processed in ~10 hrs.
- High quality variant calls-PG/CFI pipeline

Total Variants	23Me	UCLA	PG/CFI
EdgeBio	39116	25552	36458
23Me		30182	43417
UCLA			25378

% Discordance	23Me	UCLA	PG/CFI
EdgeBio	0.36%	0.39%	0.27%
23Me		0.31%	0.35%
UCLA			0.13%

Workflow for 2nd Tier Newborn Screening

Analysis & Interpretation

(Annotation, interpretation & reporting)

1h

- Tested two known cases: MSUD and PA
- Used CRADD for FP/FN testing
- Tested blinded samples, a set of 8 from 30

NO	DISORDER	POPULATION	GENE	VARIANT
1	Phenylketonuria	Amish/Mennonite	PAH	782 G>A
2	Maple Syrup Urine Disease	Mennonite	BCKDHA	1312 T>A
3	Medium Chain Acyl-Co Dehydrogenase Deficiency	Mennonite	ACADM	985 A>G
4	Glutaric Aciduria Type I	Amish	GCDH	1262 C>T
5	Cystic Fibrosis	Amish/Mennonite	CFTR	1522-24 del TTT
6	Severe Combined Immunodeficiency Disease	Amish	RAG1	2974 A>G
7	Severe Combined Immunodeficiency Disease	Mennonite	IL7R	2 T>G
8	21-Hydroxylase Deficiency	Amish	CYP21A2	518 T>A
9	11-β-Hydroxylase Deficiency	Amish	CYP11B1	1343 G>A
10	3-β-Hydroxy Steroid Dehydrogenase Deficiency	Amish	HSD3B2	35 G>A
11	Galactose Uridyl Transferase Deficiency	Amish	GALT	563 A>G
12	Biotinidase Deficiency	Mennonite	BTD	1459 T>C
13	Biotinidase Deficiency	Amish	BTD	1368 A>C
14	Biotinidase Deficiency	Amish	BTD	1330 G>C
15	Homocystinuria	Amish	MTHFR	1129 C>T
16	Propionic Acidemia B	Amish/Mennonite	PCCB	1606 A>G
17	Adenosine Deaminase Deficiency (SCID)	Amish	ADA	646 G>A
18	Glutaric Aciduria Type 3	Amish	C7orf10	895 C>T
19	3-Methylcrotonylglycinuria 2β	Amish	MCCC2	295 G>C
20	3-Methylcrotonylglycinuria 2β	Mennonite	MCCC2	518 ins T
21	3-Methylcrotonylglycinuria 2β	Mennonite	MCCC2	687 A>C
22	Mevalonate Kinase Deficiency	Mennonite	MVK	803 T>C
23	Mevalonate Kinase Deficiency	Mennonite	MVK	1174 G>A
24	Galactose Uridyl Transferase Deficiency	Amish	GALT	940 A>G
25	Phenylketonuria	Amish	PAH	280-282 del ATC
26	Phenylketonuria	Mennonite	PAH	IVS 10-11 G>A
27	Phenylketonuria	Mennonite	PAH	IVS 12+1 G>A
28	Tyrosinemia Type 3 (Hawkinsinuria)	Mennonite	HPD	85 G>A
29	Tyrosinemia Type 3 (Hawkinsinuria)	Mennonite	HPD	479 A>G
30	Tyrosinemia Type 3 (Hawkinsinuria)	Mennonite	HPD	1005 C>G
31	Methylmalonic/Homocystinuria, cblC Deficiency	Amish	MMACHC	271 ins A
32	Medium Chain Acyl-CoA Dehydrogenase Deficiency	Mennonite	ACADM	IVS 4-30 A>G

← Known

← Known

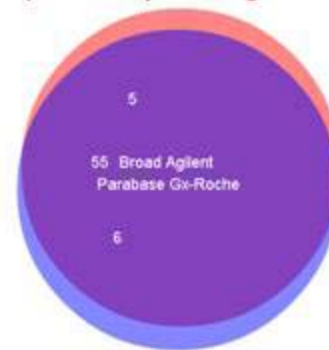
Detecting Known Cases

Identifying causal variants using population frequency and disease category as filters

Type	Sample	Disease	Exome Variants	Protein Impact (PI)	PI+ Probably Damaging	PI+PD Hom.Re ad>4 MAF <5%	I26 Gene Panel Read >4 MAF<5%	I26 GP,Comm on Hom., Read >4 MAF<5%	Gene	Reads	Transcript Variant	Protein Variant	Status
Amish*	whole blood	Propionic Acidemia	71,261	10,127	1,013	11	19	2	PCCB	18	c.1606A>G	p.Asn536Asp	Hom.
Amish*	DNA	Propionic Acidemia	64,003	10,451	1,037	15	16	2	PCCB	5	c.1606A>G	p.Asn536Asp	Hom.
Mennonite*	DNA	Maple Syrup Urine Disease	68,703	10,217	1,031	19	19	3	BCKDHA	35	c.1312 T>A	p.Tyr438Asn	Hom.
Mennonite*	DNA	Mental Retardation NS	69,946	10,329	1,086	21	ND	ND	CRADD	15	c.382G>C	p.Gly128Arg	Hom.

Validation of False Positive/ False Negatives by comparing different Methods

CRADD Sample, Broad**	Exome Variants	Panel (<i>in silico</i>)
SYNONYMOUS	11219	81
MISSENSE	10518	65
NONSENSE	91	0
SMALL INDELS	889	4
INTRON, UTRs	26083	232
SPLICE SITE	160	1



Data generated on Nimblegen Exome; variant calling GATK (Broad Institute, MA); Omicia (Emeryville, CA)

*Samples provided by Clinic for Special Children

** Puffenberger *et al.*, 2012. PLoS ONE 7(1): e28936. Agilent Exome using Broad Pipeline

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Detecting 8 blinded cases among 30

NO	DISORDER	POPULATION	GENE	VARIANT	Sample
1	Phenylketonuria	Amish/Mennonite	PAH	782 G>A	S10
2	Maple Syrup Urine Disease	Mennonite	BCKDHA	1312 T>A ←	Known
3	Medium Chain Acyl-Co Dehydrogenase Deficiency	Mennonite	ACADM	985 A>G	S1
5	Cystic Fibrosis	Amish/Mennonite	CFTR	1522-24 del TTT	S3
7	Severe Combined Immunodeficiency Disease	Mennonite	IL7R	2 T>G	S6
9	11-β-Hydroxylase Deficiency	Amish	CYP11B1	1343 G>A	S9
11	Galactose Uridyl Transferase Deficiency	Amish	GALT	563 A>G	S5
12	Biotinidase Deficiency	Mennonite	BTD	1459 T>C	S7
15	Homocystinuria	Amish	MTHFR	1129 C>T	S4
16	Propionic Acidemia B	Amish/Mennonite	PCCB	1606 A>G ←	Known

All Samples 8/8 were correctly identified

No.	Exome Variants	Protein Impact (PI) PG/CFI	PI RTG	%SNV >40	Avg. Reads	PI+PD >5			126 tal Panel	552 Here dietary panel	Gene	Reads	Transcript variant	Protein Variant	Zyg.
						PI+PD <5% AF	PI+PD <5% AF Hom	PI+PD <5% AF Het.							
S1	43373	13909	10468	95	102	10	2	10	1	4	ACADM	101	c.1085 A>G	p.Lys362Glu	Hom.
S3	43537	14142	13549	95	72	12	0	12	0(4)	3(16)	CFTR	43	c.1521_1523del	p.Ser507del	Hom.
S4	43033	13968	10314	95	89	6	1	5	2	3	MTHFR	92	c.1251C>T	p.Arg417Cys	Hom.
S5	42985	14123	10417	94	88	14	2	12	1	4	GALT	79	c.564A>G	p.Gln188Arg	Hom.
S6	43759	14080	10585	95	99	10	0	10	0(3)	0(24)	IL7R	198	c.3T>G	p.met1Arg	Hom.
S7	43706	14269	10749	95	97	14	0	14	0(4)	0(14)	BTD	74	c.1460T>C	p.Trp487Arg	Hom.
S9	43244	14051	10580	96	108	7	3	4	2	4	CYP11B1	57	c.1343G>A	p.Arg448His	Hom.
											ADA	66	c.646G>A	p.Gly216Arg	Het.
											CDH23	64	c.3309A>G	p.Asn1103Ser	Het.
S10	43356	14363	10266	92	48	16	0	16	4	15(61)	PAH	33	c.782G>A	p.Arg261Gln	Het.
											PAH	35	c.284_286del	p.(Ile95_Lys96del)	Het.
											MCC2	61	c.295G>C	p.Glu99Gln	Het.

Disease categories, allele frequency (AF) filters can quickly identify causal variants
 Panels (*in silico*) based on neonatal diseases & symptoms can rapidly identify mutants
 We were able to identify compound heterozygote (PAH) and carriers

Data was generated on variant calling of CFI and RTG and reviewed in Omicia Opal (Emeryville, CA)

•All Samples were provided by Clinic for Special Children

•Variant identification was faster on *in silico* neonatal panels than on entire Exome

Summary

- Can be used as 2nd Tier Newborn testing including NICU
- Causal variants identified rapidly in targeted panels
- Targeted NGS panel can be used for screening monogenic disorders
- Targeted NGS is affordable unlike Whole Genome (\$\$\$)
- Targeted NGS is efficient- Do >10 'samples to answer' in 100 Hrs.

For product, services and collaboration opportunities please contact us

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Sample Isolation

Parabase Genomics

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Thank You